

Preliminary study of marine sponges (Porifera) in the littoral of Spermonde Archipelago, Indonesia

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Abstract

Previous ecological studies show higher sponge diversity in the Spermonde Archipelago, SW Sulawesi, Indonesia, compared to the World Porifera Database. This study aims to provide an updated checklist of sponges of the Spermonde Archipelago, focusing particularly on the littoral area. Systematic sampling was executed through several observations, with roving techniques, e.g., snorkeling and SCUBA diving. In situ photographs of living sponges were taken using an underwater digital camera. Some specimens were collected and stored at the Naturalis Biodiversity Center, Leiden. Fragments of samples were analyzed using light and scanning electron microscopy. A total of 27 sponges (Calcarea and Demospongiae) were catalogued from the littoral area of the Spermonde Archipelago. Some of these are new records for the Sulawesi Sea/Makassar Strait marine ecoregion, including four potentially novel taxa. Preliminary morphological descriptions of all examined samples are presented. This study highlights the sponge assemblage flourishing in a shallow area characterized by a paucity of live corals and a predominant environment by macroalgae, rocks, and rubble.

Key words: Calcarea, Demospongiae, Indo-Pacific, taxonomy, turbid habitats



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Introduction

The Spermonde Archipelago is located between the south-western part of Sulawesi and the Makassar Strait in Indonesia (Kench and Mann 2017). This region is placed in the Sulawesi Sea/Makassar Strait (SS/MS) marine ecoregion based on Marine Ecoregions of the World (Spalding et al. 2007). The whole archipelago consists of many coral cays and small islands (Umbgrove 1928; Kench and Mann 2017), with the highest coral cover less than 60% (Sari et al. 2021). The coral reef is the richest ecosystem with high species diversity (Cairns 1999; Williams et al. 2019; Kusumoto et al. 2020). Every part of the reef is influenced by different regimes of wave actions, light intensity, bathymetric range, and water currents (Kench and Mann 2017). The sponge community

is one of the essential components of the reef environment (Rützler 2004), showing a wide distribution across the Spermonde Archipelago (de Voogd et al. 1999). This community is also recognized as comprising predominantly niche specialists with marked habitat preferences in coral reef ecosystems (Hooper 2008).

Numerous studies have been conducted on this archipelago due to its geological, biodiversity, and ecological significance in marine biology (Polónia et al. 2015). Taxonomic studies on sponge diversity in this region were sporadic. The sponge fauna within the SS/MS marine ecoregion is relatively well studied only in north Sulawesi (de Voogd et al. 2024). Only a few papers have conducted morphological taxonomic studies to describe new species or revise specific group of sponges (e.g., genus, family, or order), with a mention of the Spermonde Archipelago as a locality (de Weerd and van Soest 2001; de Voogd 2004; Becking 2013; Alvarez et al. 2016; van Soest et al. 2021).

Globally, more than 9,000 sponge species are currently described (de Voogd et al. 2024). Taxonomic misidentifications by non-taxonomists are common when dealing with sponges (Cárdenas et al. 2022). Some comprehensive inventories of the sponge fauna from Indonesia have been published (van Soest 1990; Calcinaï et al. 2017), including specific sponge category-based inventories (de Voogd and van Soest 2002; Calcinaï et al. 2005; van Soest and de Voogd 2015; van Soest et al. 2021). However, sponge diversity across the Indonesian Archipelago is still considered underestimated (Calcinaï et al. 2017; Putra et al. 2023).

According to the World Porifera Database (de Voogd et al. 2024), sponge diversity in the Sulawesi Sea/Makassar Strait marine ecoregion comprises 128 species, i.e., 17 species of Calcarea, 97 species of Demospongiae, 13 species of Hexactinellida, and one species of Homoscleromorpha. The class Demospongiae is predominantly represented by the order Poecilosclerida, comprising 31 species. However, the latest ecological study reveals a higher sponge species beta diversity in the Spermonde Archipelago, SW Sulawesi. At least 151 species belonging to 68 genera and 37 families were identified in this area (de Voogd et al. 2006). Therefore, taxonomic studies are needed to describe the unregistered sponge species and elucidate the sponge alpha diversity in this marine ecoregion.

The current study is focused on the littoral area of the archipelago. This area is below the lowest tide, but including the reef flat. Reef flats are the most recent expression of sea-level coral reef growth (Hopley 2011). This area presents extreme conditions for coral reefs due to marginal environmental factors (Burt et al. 2020). Furthermore, the coral reef ecosystem in this shallow area, particularly in the inner zone of the archipelago, was reported to be in a very poor condition, $\approx 5\text{--}14\%$ (Paren den et al. 2021; Sari et al. 2021). This habitat is dominated by dead corals with algae, macroalgae, and sediment cover (Paren den et al. 2021).

This study aims to provide preliminary morphological identifications of sponge specimens from the Spermonde Archipelago to fill the knowledge gap concerning marine sponge diversity of Indonesia. Additionally, it seeks to promote the study of sponge taxonomy in Indonesia and to update the checklist of sponge diversity of this marine ecoregion.

Materials and methods

Specimen collection

The specimen collection was conducted through several observations of the littoral area of the Spermonde Archipelago, Indonesia. Some observations were made by NJdeV in 2018, and by SAP during 2020 and 2021 (Fig. 1, Suppl. material 1). The observations were performed using a roving technique (Pattengill-Semmens 2001) through snorkeling or SCUBA Diving. Roving time is 1–2 hours within $\approx 90 \text{ m}^2$ for each site. The timed survey method does not provide density and abundance data but is most useful when the study aims to assess biodiversity (Reimer et al. 2018; Montano et al. 2020).

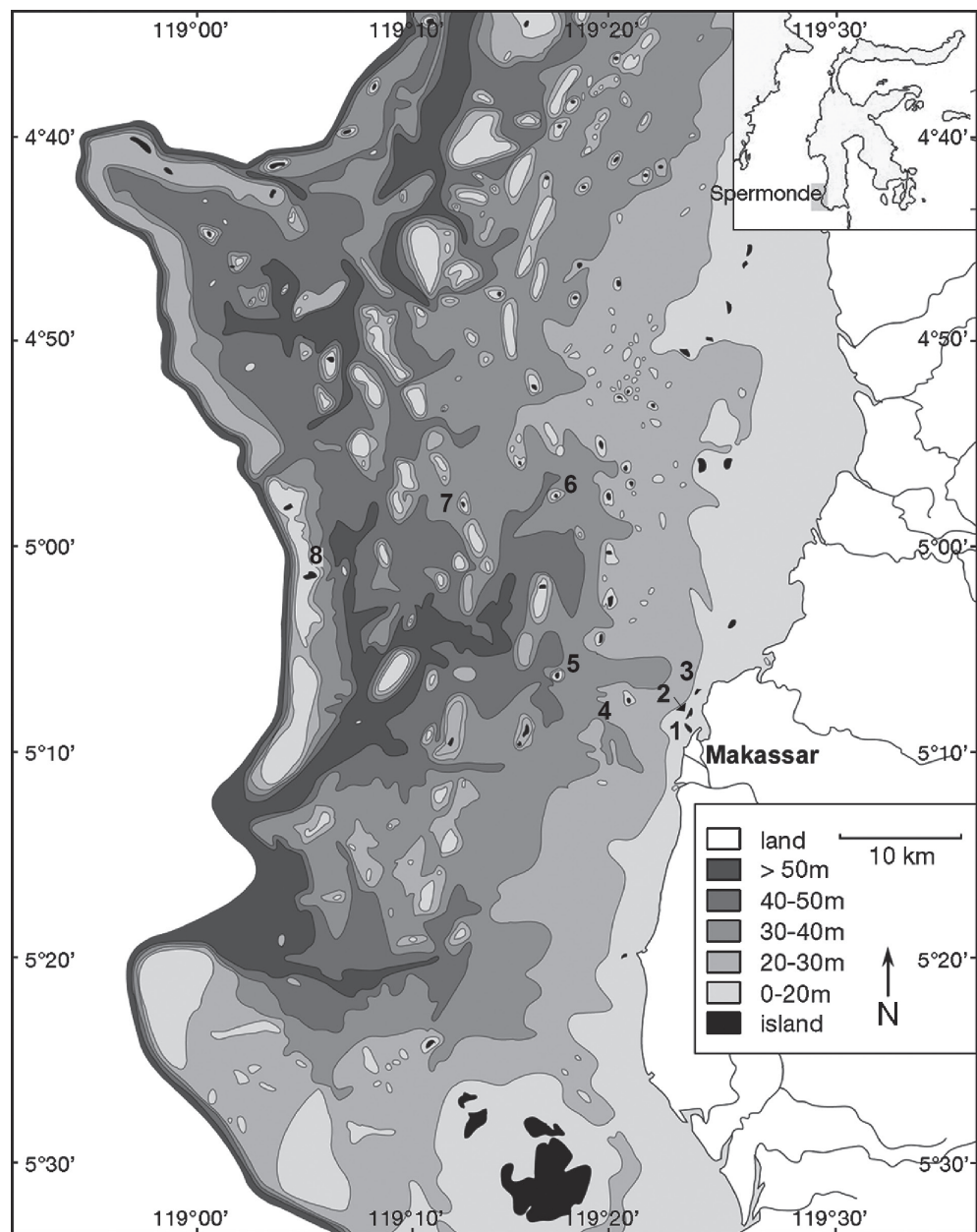


Figure 1. The location of sponge observation in the shallow-subtidal area of the Spermonde Archipelago, SW Sulawesi, Indonesia, i.e., 1) Lae-lae, 2) Gusung (as Gusung Tallang), 3) Kayangan, 4) Samalona, 5) Kudingarengkeke, 6) Badi, 7) Lumulumu, 8) Langkai.

Photographs of living sponges at the study site (in situ) were captured using an underwater digital camera (Nikon Coolpix W300 and Olympus TG-series). The specimens were immediately transferred into 96% ethyl alcohol for preservation during observation (Hooper 2003), and some of them were deposited in the museum collection of the Naturalis Biodiversity Center, Leiden, The Netherlands (**NBC**); the others are located at Balai Pengembangan Penjaminan Mutu Pendidikan Vokasi Bidang Kelautan Perikanan Teknologi Informasi dan Komunikasi (**BPPMPV KPTK**) in Gowa, Sulawesi Selatan.

Specimen identification

Fragments of sponges and sections of the skeleton were prepared and then examined using light microscopy (Leica DM5500 B and Olympus BX53) and JEOL Scanning Electron Microscope (JSM-6480LV) at the Naturalis Biodiversity Center, Leiden, following standard procedures for skeleton and spicule analysis (Rützler 1974; Boury-Esnault and Rützler 1997; Hooper 2003). Except for macro morphologies, which were measured with a vernier caliper, microscopic characteristics were assessed using Olympus cellSens Standard and Leica LAS Core software. Images were cleaned up and assembled in composite figures using Adobe Photoshop 2023 and Adobe Illustrator 2023 licensed to SAP. Measurements of spicules (smallest-largest-(mean)) rely on a minimum of 20 measurements of length and thickness for each type of spicule in the case of one or a few specimens. Systematic treatment refers to the description of Porifera morphological identification (Hooper and van Soest 2002) and the World Porifera Database/WPD (de Voogd et al. 2024). The recording of species names includes as much information as possible, such as valid names, species location, specimen description, and other taxonomic notes.

Results

Systematics

Accepted names, all synonyms, and systematic updates were based on the World Porifera Database (de Voogd et al. 2024), and all terminology follows updated terms (Boury-Esnault and Rützler 1997; Lukowiak et al. 2022).

Phylum Porifera Grant, 1835

Class Calcarea Bowerbank, 1862

Subclass Calcinea Bidder, 1898

Order Clathrinida Hartman, 1958

Family Clathrinidae Minchin, 1900

Genus *Clathrina* Gray, 1867

***Clathrina rodriguesensis* van Soest & de Voogd, 2018**

Fig. 2

Diagnostic features. In its natural environment, the species forms a large, encrusting mass composed of wide, closely linked tubes showing little variation in diameter. According to van Soest and de Voogd (2018), it can

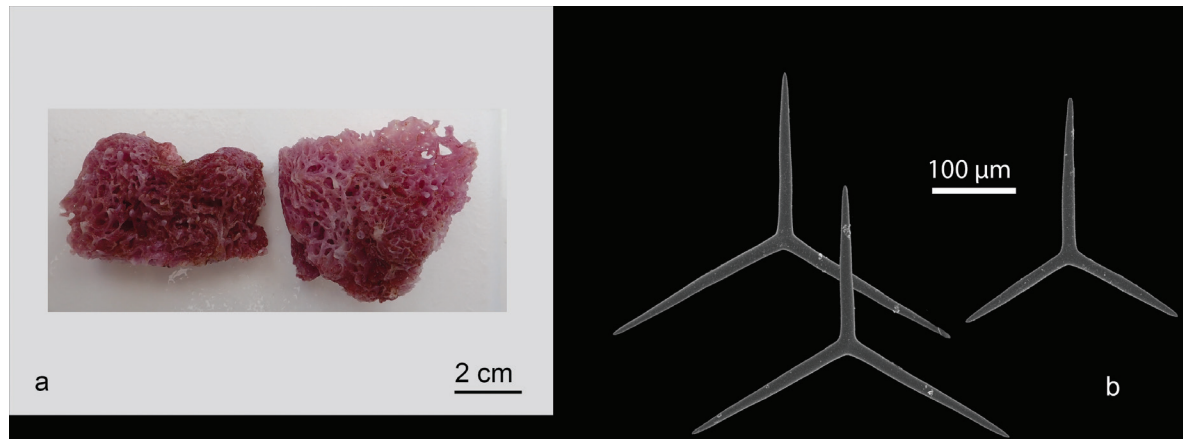


Figure 2. *Clathrina rodriguesensis* van Soest & de Voogd, 2018 from Kudingareng keke Island, the Spermonde Archipelago (Sample CEL035) **a** habitus of fresh specimen (photograph by NJdeV) **b** SEM image of the triactines.

spread flatly across wide areas, with the tubes arranged like a ladder. The main tubes often end in an opening slightly raised from the mass. The color is white with shades of blue, grey, or pink, turning pale beige or brown when preserved. Consistency firm and with asconoid aquiferous system. Spicules are only triactines.

Distribution and ecology. Previously, this species only recorded from Seychelles, Western Indian Ocean (van Soest and de Voogd 2018). This is first record for Indonesia (Kudingareng keke, the Spermonde Archipelago; reef flat).

Genus *Janusya* Klautau et al., 2021

Janusya tubuloreticulosa (van Soest & de Voogd, 2015)

Fig. 3

Diagnostic features. An orange flattened mass of short oscular tubes, connected at the substratum by a basal tubular network, the erect tubes maybe divided into one or two side tubes. The walls of tubes are thin with spicules are dominated by triactines. Triactines predominantly equiactinal with size $14.93\text{--}120.79$ (83.54) \times $3.39\text{--}6.76$ (5.48) μm ($n = 20$). Tetractines are also not rarely found with size $28.04\text{--}103.79$ (83.77) \times $4.98\text{--}5.94$ (5.48) μm ($n = 11$).

Distribution and ecology. Originally reported from Ternate (van Soest and de Voogd 2015). First record from Samalona Island, the Spermonde Archipelago; reef flat.

Family Leucaltidae Dendy & Row, 1913

Genus *Laucaltis* Haeckel, 1872

Leucaltis nodusgordii (Poléjaeff, 1883)

Fig. 4

Diagnostic features. The species forms a clathrate mass of interconnected (anastomosing) tubes with varying lengths and diameters. Individual tubes can reach up to 2.5 cm in length and have diameters of 2–8 mm (van Soest and de Voogd 2015). The tubes end in oscula, which can be as wide as the

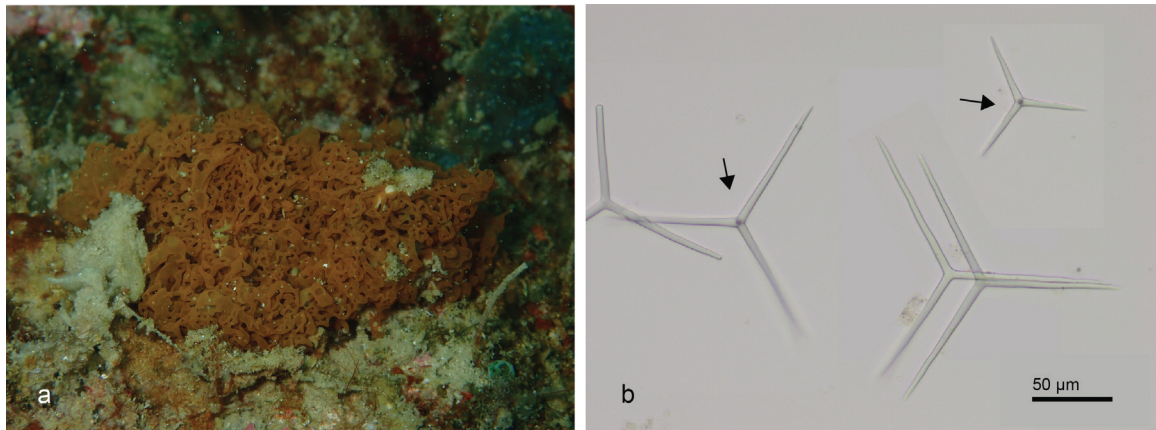


Figure 3. *Janusya tubuloreticulosa* (van Soest & de Voogd, 2015) from Samalona Island, the Spermonde Archipelago (Sample CEL001) **a** habitus in situ at Samalona reefs (photograph by NJdeV) **b** LM images of spicules, triactines and tetractines (arrows).

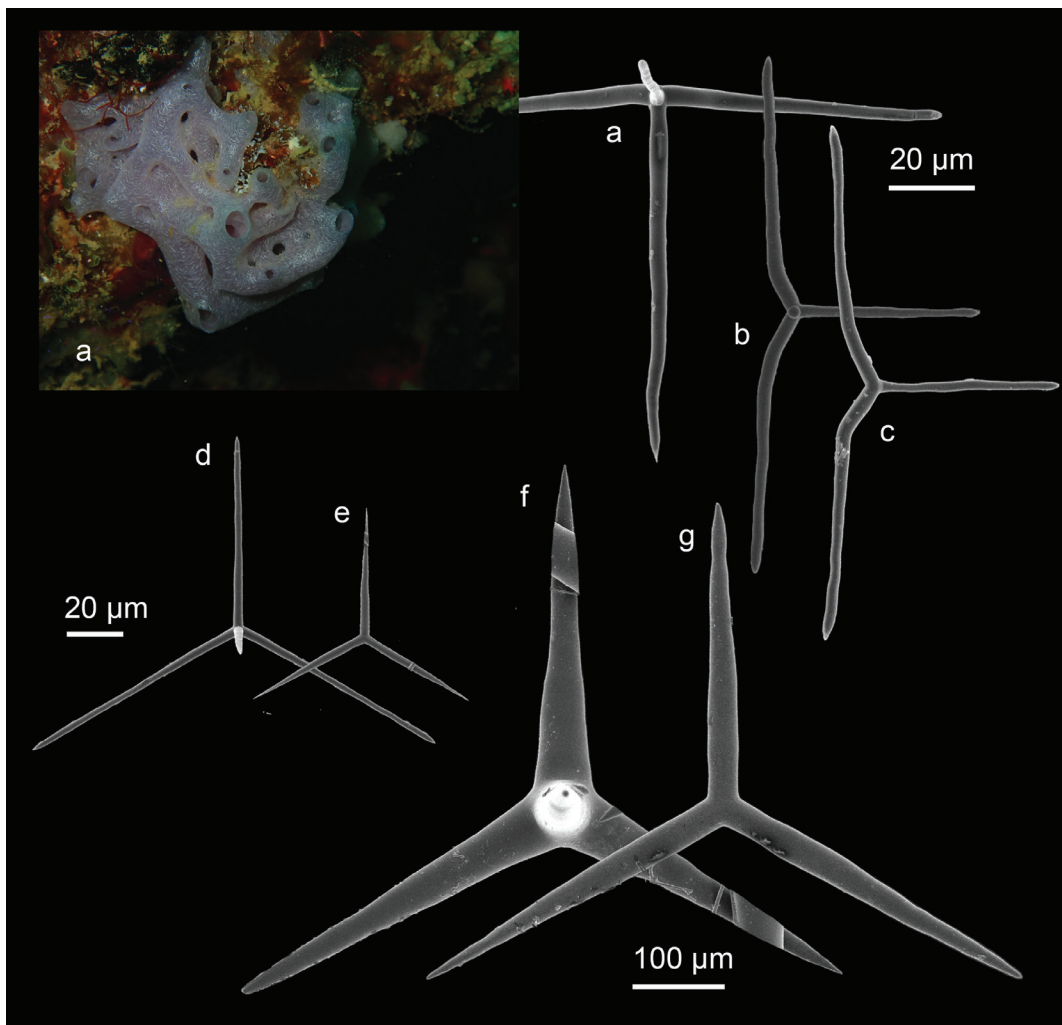


Figure 4. **a** habitus in situ *Leucaltis nodusgordii* (Poléjaeff, 1883) (CEL005) from Samalona Island, the Spermonde Archipelago (photograph by NJdeV). SEM images of spicules **a** regular equiangular tetractine of the chamber layer **b** 'Abruptly angled' tetractines **c** 'Abruptly angled' triactines (**b**, **c** both from the atrial region) **d** Small regular-shaped tetractines of the chamber layer **e** small regular-shaped triactines of the cortical region **f** Giant sized tetractines **g** giant sized triactines (**f**, **g** both from the cortical region).

tube itself (standing upright) or smaller (flush with the surface), and these oscula are naked. The surface is smooth, and the texture is brittle yet somewhat compressible. The color is white or pinkish white, sometimes lavender-colored, and it turns yellowish white when preserved. The cortical skeleton is formed by the basal triradiate system of giant tetractines mixed with giant triactines. Actines of the giant tetractines and triactines protrude into the choanosomal skeleton. Next to the actines of the giant tri- and tetractines, the choanosomal skeleton contains scattered intermediate to small-sized regular triactines and tetractines (see van Soest and de Voogd 2015 for detail description).

Distribution and ecology. *Leucaltis nodusgordii* is a new record for the Spermonde Archipelago (Samalona Island); reef flat. This species has been reported previously from north Sulawesi (van Soest and de Voogd 2015).

Class Demospongiae Sollas, 1885

Subclass Heteroscleromorpha Cárdenas et al., 2012

Order Clionaida Morrow & Cárdenas, 2015

Family Spirastrellidae Ridley & Dendy, 1886

Genus *Spirastrella* Schmidt, 1868

***Spirastrella* aff. *decumbens* Ridley, 1884**

Fig. 5

Diagnostic features. A thin encrusting sponge with a soft texture and a smooth surface. The living specimens exhibit a salmon-pink or orangish color. The ectosome of the sponge contains numerous microscleres (spirasters), forming the characteristic tangential crust found in this genus. In the choanosome, the megascleres are irregularly arranged tylostyles with well-formed, usually spherical heads (Calcinai et al. 2006). Our specimen shows spirasters with ornamented rays (Fig. 5d) that are not mentioned in the Calcinai et al. (2006) report from Vietnam.

Distribution and ecology. This species is present in the Australian region, New Caledonia, the Philippines, and Vietnam. In Indonesia is recorded from Ambon; this is a first record for the Spermonde Archipelago (Langkai Island; reef flat).

Order Haplosclerida Topsent, 1928

Family Callyspongiidae de Laubenfels, 1936

Genus *Callyspongia* Duchassaing & Michelotti, 1864

Subgenus *Cladochalina* Schmidt, 1870

***Callyspongia* (*Cladochalina*) *johannesthielei* van Soest & Hooper, 2020**

Fig. 6

Diagnostic features. Lobate form and hard surface with numerous, raised, cone-shaped projections (pointed papillae). Several large oscula between \approx 6–7 mm. Pink to red in living and pale yellow in alcohol. The skeleton is reticulate with a fiber tract. This species was described as *Spinossella elegans* Thiele, 1899 (junior secondary homonym of *Callyspongia* (*Cladochalina*) *ele-*

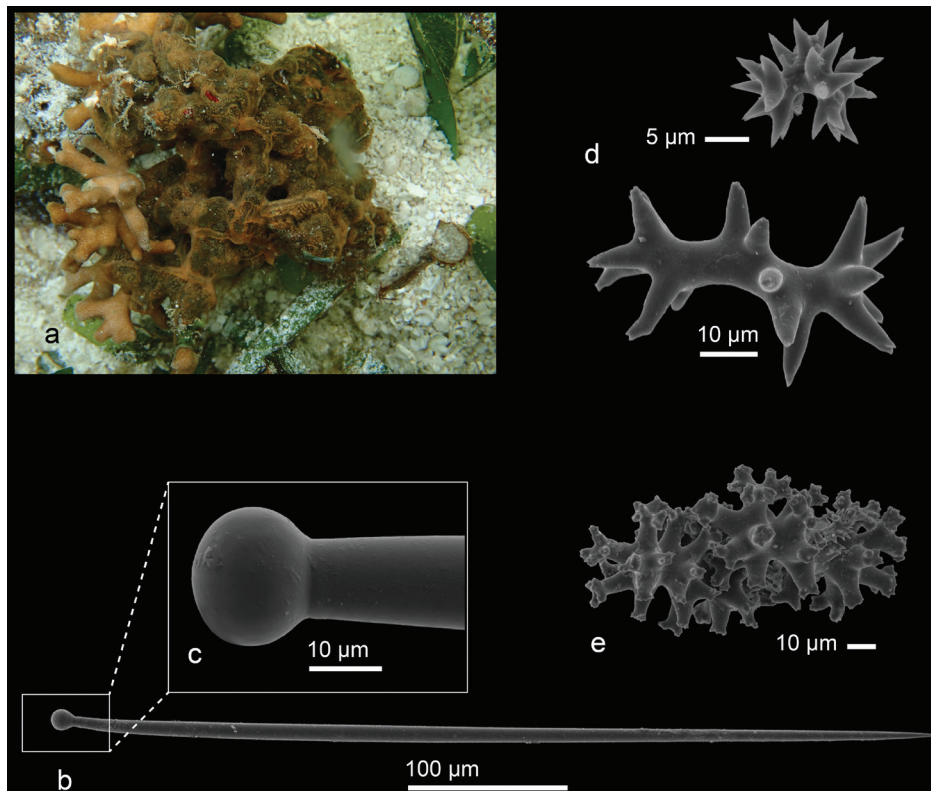


Figure 5. *Spirastrella* aff. *decumbens* Ridley, 1884, overgrowing coral skeleton **a** habitus in situ (CEL007) from seagrass bed of Langkai Island, the Spermonde Archipelago (photograph by NJdeV) **b** SEM image of tylostyle with **c** close up of the head **d** spirasters **e** spirasters with ornamented rays.

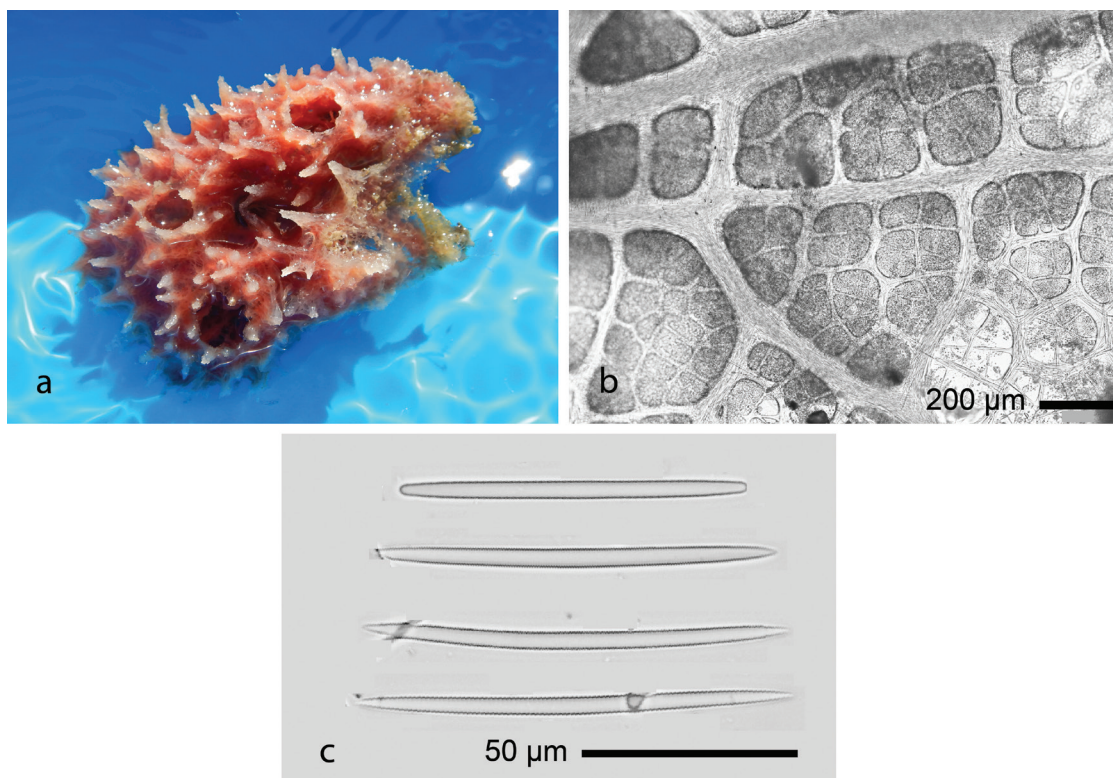


Figure 6. *Callyspongia* (*Cladochalina*) *johannesthielei* van Soest & Hooper, 2020 **a** habitus of fresh specimen (photograph by SAP) **b** skeleton **c** amphioxeas.

gans (von Lendenfeld, 1887)) as a large cup-shaped sponge, ≈ 30 cm high, hollow along its entire length, a pale brownish color when dry, and with very characteristic pointed papillae, often fused into a cluster of several, on the outer surface (Thiele 1899). The spicules of Thiele's species were shown as rather thin, short-tipped amphioxeas that are $90\text{--}100\ \mu\text{m} \times 3\text{--}5\ \mu\text{m}$ (van Soest et al. 2020).

Distribution and ecology. Kema Bay ($1^{\circ}23'N$, $125^{\circ}04'E$), north Sulawesi (Thiele 1899); and north-west of Samalona Island, the Spermonde Archipelago; reef flat; attached on rock.

Family Chalinidae Gray, 1867

Genus *Haliclona* Grant, 1841

Subgenus *Gellius* Gray, 1867

***Haliclona* (*Gellius*) *cymaeformis* (Esper, 1806)**

Fig. 7

Diagnostic features. The appearance is thickly encrusting to repent or arborescent (bushy). The specimen is hard and smooth on the surface, with a broad erect base with short branches. The color in life is dark greyish pink (dark purple) with desaturated dark green on the tips. After preservation, the color is pale pink to yellow. Ectosomal skeleton shows unispicular tract and covering the associated branching microalgae (Fig. 7c). Spicules are oxeas, $109\text{--}154$ (129.7) $\times 2.3\text{--}5.2$ (3.9) μm ($n = 27$), and microscleres are sigmas.

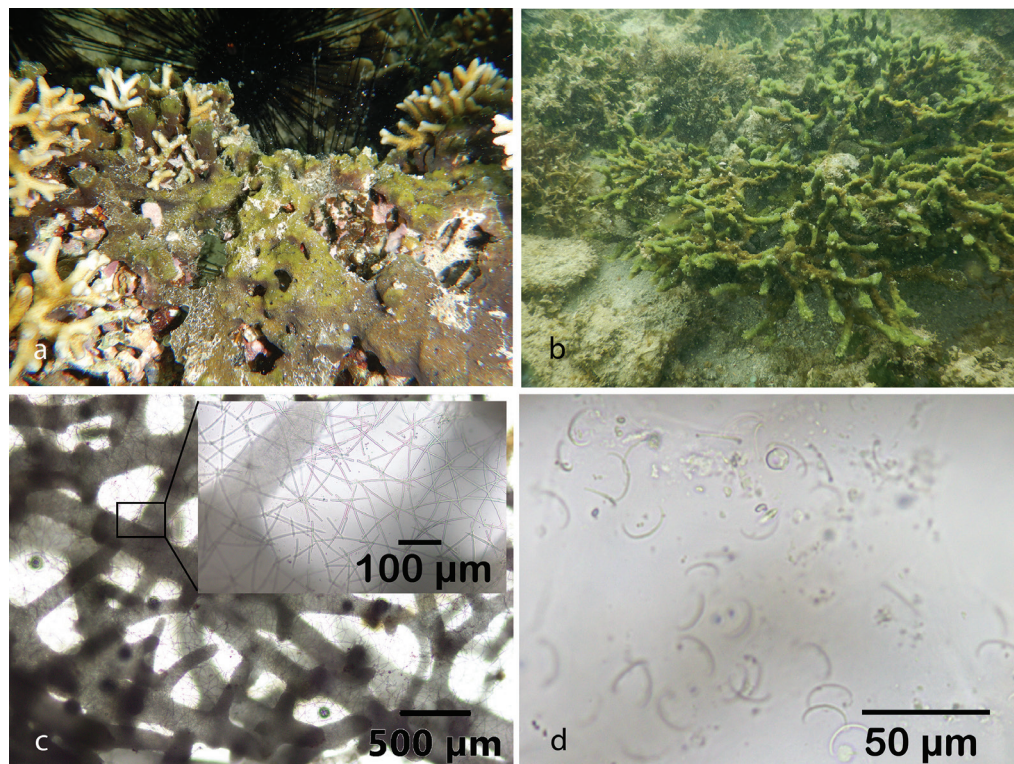


Figure 7. *Haliclona* (*Gellius*) *cymaeformis* (Esper, 1806) **a, b** habitus in situ at Samalona Island and Kayangan Island (respectively), the Spermonde Archipelago (all photographs by SAP) **c** LM images of tangential section showing Rhodophyta symbiont and unispicular tract (box) **d** sigmas.

Haliclona (Gellius) cymaeformis (Esper, 1806) was abundant in turbid water near Makassar City. This species is known to be associated with the rhodophyte *Ceratodictyon spongiosum* Zanardini, 1878 (Azzini et al. 2007). Its morphological appearance is possibly similar to those of *Halichondria (Halichondria) cartilaginea* (Esper, 1797) and *Callyspongia (Cladochalina) samarensis* (Wilson, 1925).

Distribution and ecology. This species has been recorded from marine karst lakes in Vietnam (Azzini et al. 2007), in shallow waters of the South China Sea (Huang et al. 2016; Lim et al. 2016), and in Taiwan (Li 2013), Andaman (Immanuel et al. 2015), India (George et al. 2020), across the Indonesian Archipelago (de Voogd and Cleary 2008), and north-west Australia (Fromont and Sampey 2014). Our samples were collected from a reef flat north-west of Samalona Island, overgrowing corals (*Seriatopora* sp. and *Acropora* sp.), also from Kayangan Island and Gusung Tallang; turbid reef environment.

Subgenus *Reniera* Schmidt, 1862

Haliclona (Reniera) venusta (Bowerbank, 1875)

Fig. 8

Diagnostic features. Specimen form tube, soft and delicate. Color yellowish in living material and yellow to pale white in alcohol. The skeleton forms an isotropic reticulation of a single line spicules. All spicules on this specimen are oxeas, 88–109 (95.2) × 4.3–6.5 (5.7) μm ($n = 20$).

Distribution and ecology. The WPD checklist only lists four species of the subgenus *Reniera* recorded from marine ecoregions of Indonesia with two as doubtful species, *Haliclona (Reniera) cinerea* (Grant, 1826) (doubtful species), *Haliclona (Reniera) fascigera* (Hentschel, 1912), *Haliclona (Reniera) infundibularis* (Ridley & Dendy, 1887) (doubtful species), and *Haliclona (Reniera) venusta* (Bowerbank, 1875), but none of these species were registered in the Spermonde Archipelago (Putra et al. 2023). This report presents a new record of *Haliclona (Reniera) venusta* from the Spermonde Archipelago (Samalona Island; reef flat). Previously, this species has been only reported from Malacca Strait (Bowerbank 1875).

Subgenus *Soestella* de Weerd, 2000

Haliclona (Soestella) elegantia (Bowerbank, 1875)

Fig. 9a, d, e

Diagnostic features. Small specimen (l × w × h; 46 × 34 × 30 mm) and fragile, found growing in turbid water near the coastal city of Makassar. Massive shape with large oscula (3–4 mm in diameter). Color in life deep blue and pale white in alcohol. The choanosomal skeleton is paucispicular tracts. Spicules are oxeas, larger oxeas 163.9–196.2 (163.9) × 7–9.9 (8) μm ($n = 20$) and thin oxeas 92–156.1 (127.5) × 0.8–5.7 (3) μm ($n = 26$). Microscleres are sigmas. The subgenus *Haliclona (Soestella)* consists of 25 species,

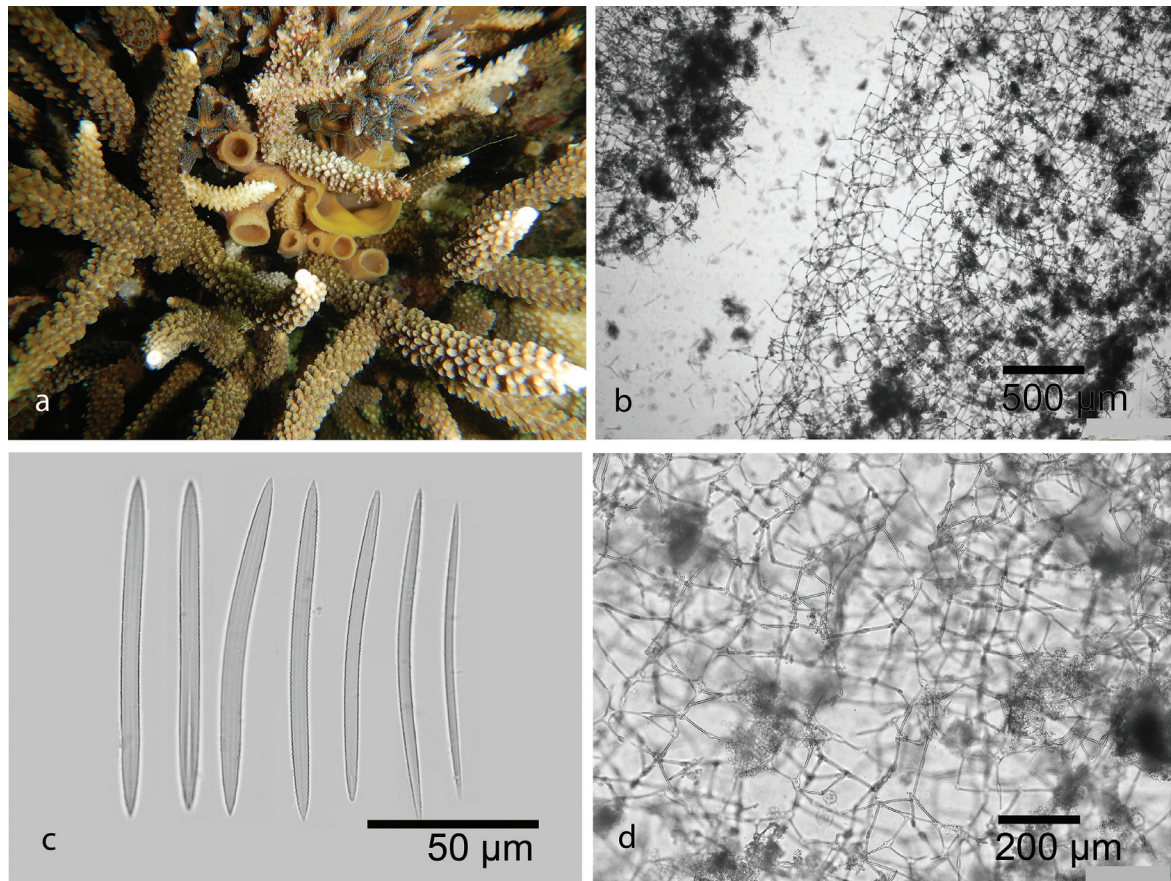


Figure 8. *Haliclona (Reniera) venusta* (Bowerbank, 1875) **a** habitus in situ at Samalona Island, the Spermonde Archipelago (photograph by SAP) **b** LM images of tangential section and spicules reticulation **c** oxeas **d** close up of spicules reticulation.

and only *Haliclona (Soestella) elegantia* is registered from the marine ecoregions of Indonesia (Putra et al. 2023). This species is poorly studied; in fact, we have found no studies after its original description. Bowerbank's description did not include an illustration, but the specimen was described as of small appearance and small spicules (short and stout) with fragile and elegant uni-, bi-, and tri-spiculose reticulation on the dermal structure (Bowerbank 1875: 286).

Distribution and ecology. Previously recorded from Malacca Strait (Bowerbank 1875). This is the first record for the Spermonde Archipelago (at Kayangan Island, and Gusung Tallang Island; turbid environment).

Haliclona (Soestella) sp. 1

Fig. 9b

Diagnostic features. The specimen is fragile and shapeless (amorphous), the surface is slick and smooth; the color in life is mostly black, also in alcohol. Oscula present with 1–3 mm diameter. The spicule arrangements are oxeas 101–162 (128.8) × 1.5–7 (4.9) µm ($n = 21$).

Distribution and ecology. North-west Samalona Island, the Spermonde Archipelago; reef flat.

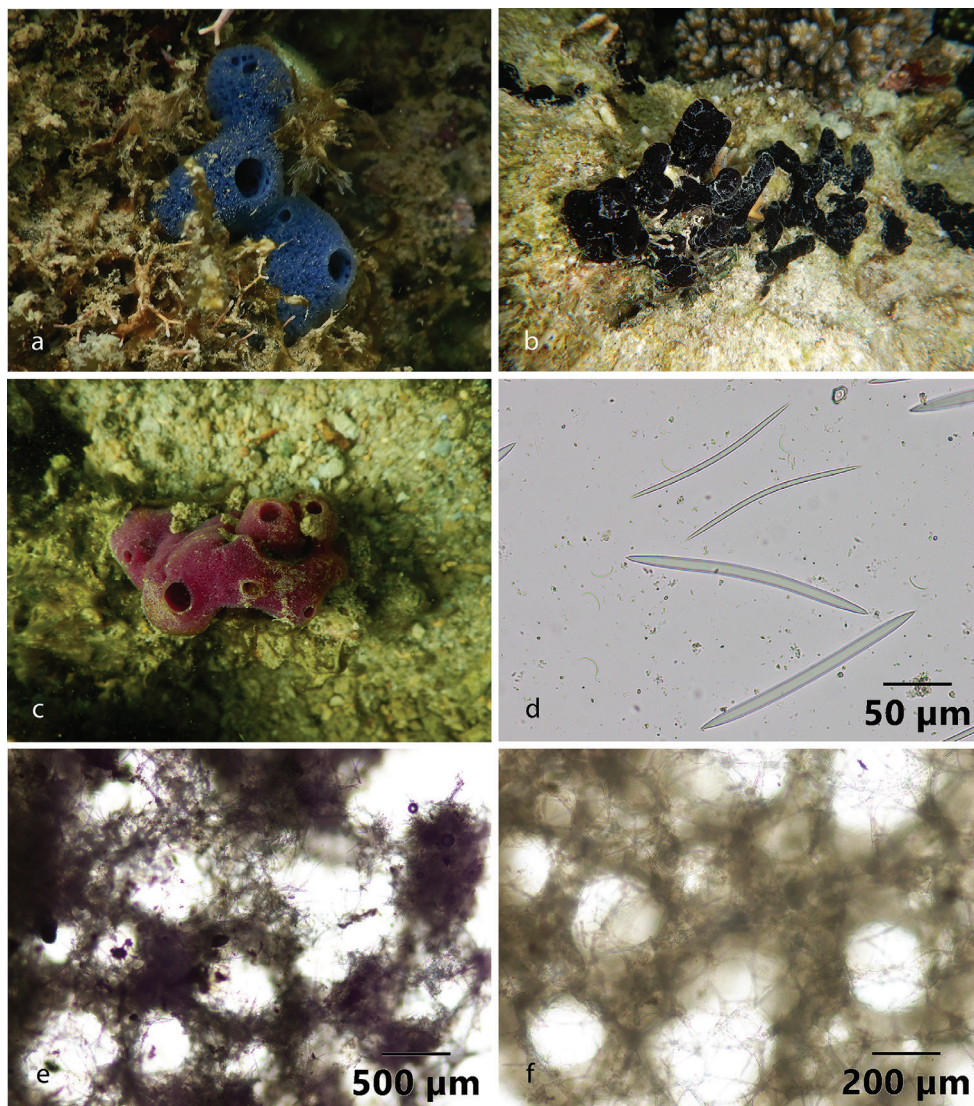


Figure 9. Habitus in situ **a** *Haliclona (Soestella) elegantia* (Bowerbank, 1875) at Kayangan Island, the Spermonde Archipelago **b** *Haliclona (Soestella)* sp. 1. at Samalona Island, the Spermonde Archipelago **c** *Haliclona (Soestella)* sp. 2. at Samalona Island, the Spermonde Archipelago (all photographs by SAP) **d** two sizes of oxeas and sigmas of *Haliclona (Soestella) elegantia* **e** *Haliclona (Soestella) elegantia* spicules reticulation **f** *Haliclona (Soestella)* sp. 2. spicule reticulation.

Haliclona (Soestella) sp. 2

Fig. 9c, f

Diagnostic features. Small specimen (l × w × h; 45 × 32 × 25 mm) with magenta color in life and pale white in alcohol. Massive shape with large osculum. Ectosomal skeleton shows multispicular fiber tracts. Spicules are oxeas, larger oxeas 102–130.9 (116.1) × 3.8–6.5 (5) µm (*n* = 24), and thin oxeas 78.4–114.4 (96.8) × 1.3–4.1 (2.5) µm (*n* = 20). Rounded meshes formed by the spicules characterized those species as belonging to the subgenus *Soestella* (de Weerd 2000). However, due to differences in color and variation of the macro-morphology, it can be distinguished from *Haliclona (Soestella) elegantia*.

Distribution and ecology. West Kayangan Island and Gusung Tallang Island, the Spermonde Archipelago; turbid environment.

Family Niphatidae van Soest, 1980

Genus *Amphimedon* Duchassaing & Michelotti, 1864

***Amphimedon paravidis* Fromont, 1993**

Fig. 10

Diagnostic features. Encrusting and soft, with small oscula and scattered ostia on the surface. Pale green in life and turning brown in alcohol. Skeleton isotropic reticulation arranged by oxeas $155\text{--}194$ (173.5) \times $5.9\text{--}8.1$ (7.2) μm . *Amphimedon paravidis* has similarities with *Amphimedon viridis* Duchassaing & Michelotti, 1864 from the Caribbean Sea. However, the holotype of *A. paravidis* (from the Great Barrier Reef) has thicker spicules, a much greater spongin component, thicker fibers, and larger mesh spaces compared to *A. viridis* (Fromont 1993). Only three species of *Amphimedon* have been reported from the marine ecoregions of Indonesia (Putra et al. 2023), including *Amphimedon anastomosa* Calcinai et al., 2017, *Amphimedon zamboangae* (Lévi, 1961), and *Amphimedon denhartogi* de Voogd, 2003.

Distribution and ecology. Previously reported from Australia (Fromont 1993). This is first record of *Amphimedon paravidis* from Samalona Island, the Spermonde Archipelago. Reef flat, overgrowing another sponge, *Clathria (Thalysias) reinwardti* Vosmaer, 1880.

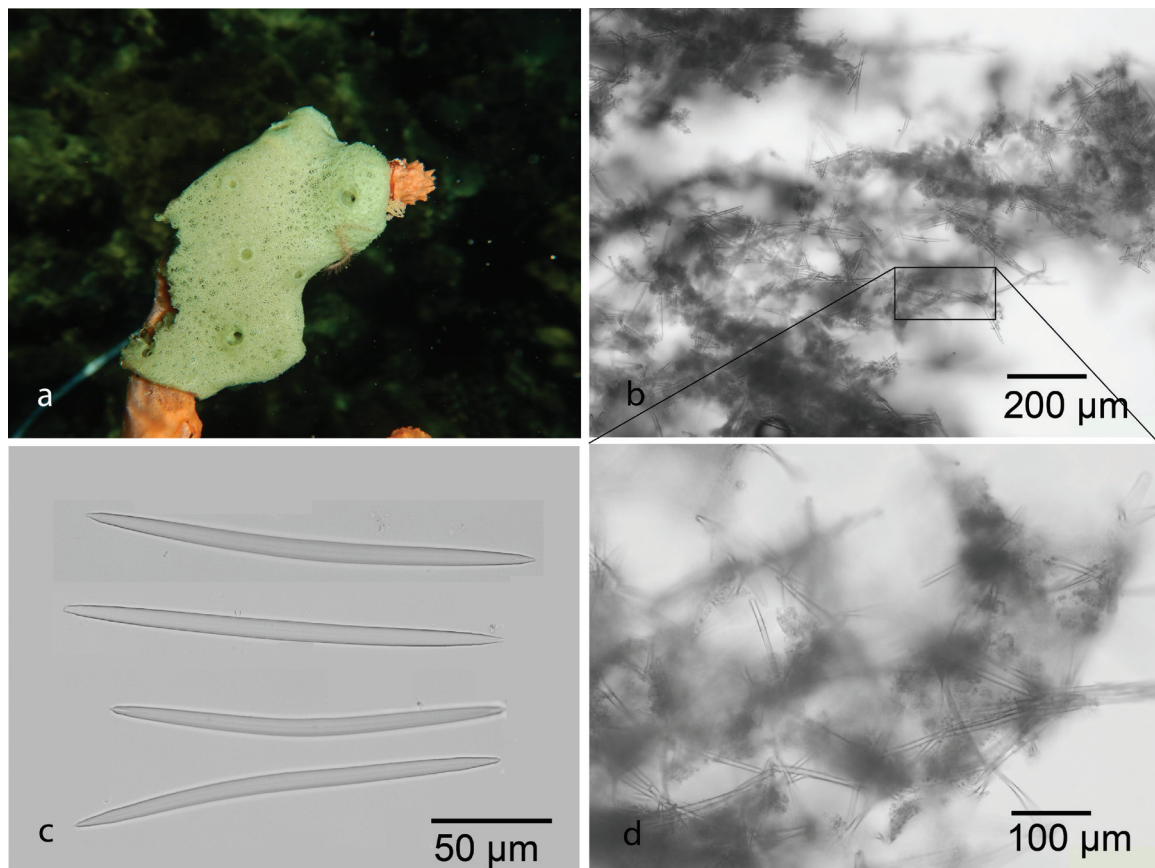


Figure 10. *Amphimedon paravidis* Fromont, 1993 **a** habitus in situ over growing *Clathria (Thalysias) reinwardti* Vosmaer, 1880 at Samalona Island, the Spermonde Archipelago (photograph by SAP) **b** LM image of cross section of the skeleton **c** oxeas **d** Isotropic reticulation of oxeas.

Genus *Niphates* Duchassaing & Michelotti, 1864

Niphates nitida Fromont, 1993

Fig. 11

Diagnostic features. Ramose repent sponge. Bluish green in life, pale white in alcohol. Oscula are small, 2–4 mm in diameter. Ectosomal shows reticulation fiber tract. Oxeas slightly curved, larger oxeas $120.3\text{--}171.3$ (139.4) \times $4.8\text{--}9.3$ (6.1) μm ($n = 22$), thin oxeas $109.3\text{--}132.7$ (121) \times $2.4\text{--}5.3$ (3.5) μm ($n = 14$). Microscleres are C-shaped sigmas. This specimen is identified as *Niphates nitida* due to the reticulation fiber tract on the skeleton and the present of sigmas. Previously, only two species of *Niphates* recorded from Indonesia. *Niphates laminaris* Calcinai et al., 2017 is characterized by a non-spiny, rather irregular, microconulose surface and a chaonosomal skeleton with primary and secondary reticulation fiber tracts, as well as numerous microscleres (Calcinai et al. 2017). *Niphates olemda* (de Laubenfels, 1954) is a tubular sponge with small oxeas (de Laubenfels 1954). *Niphates nitida* is a new record for Indonesia.

Distribution and ecology. Previously was reported from Magnetic Island, Australia (Fromont 1993). This is first record for the Spermonde Archipelago (at Kayangan Island; turbid environment).

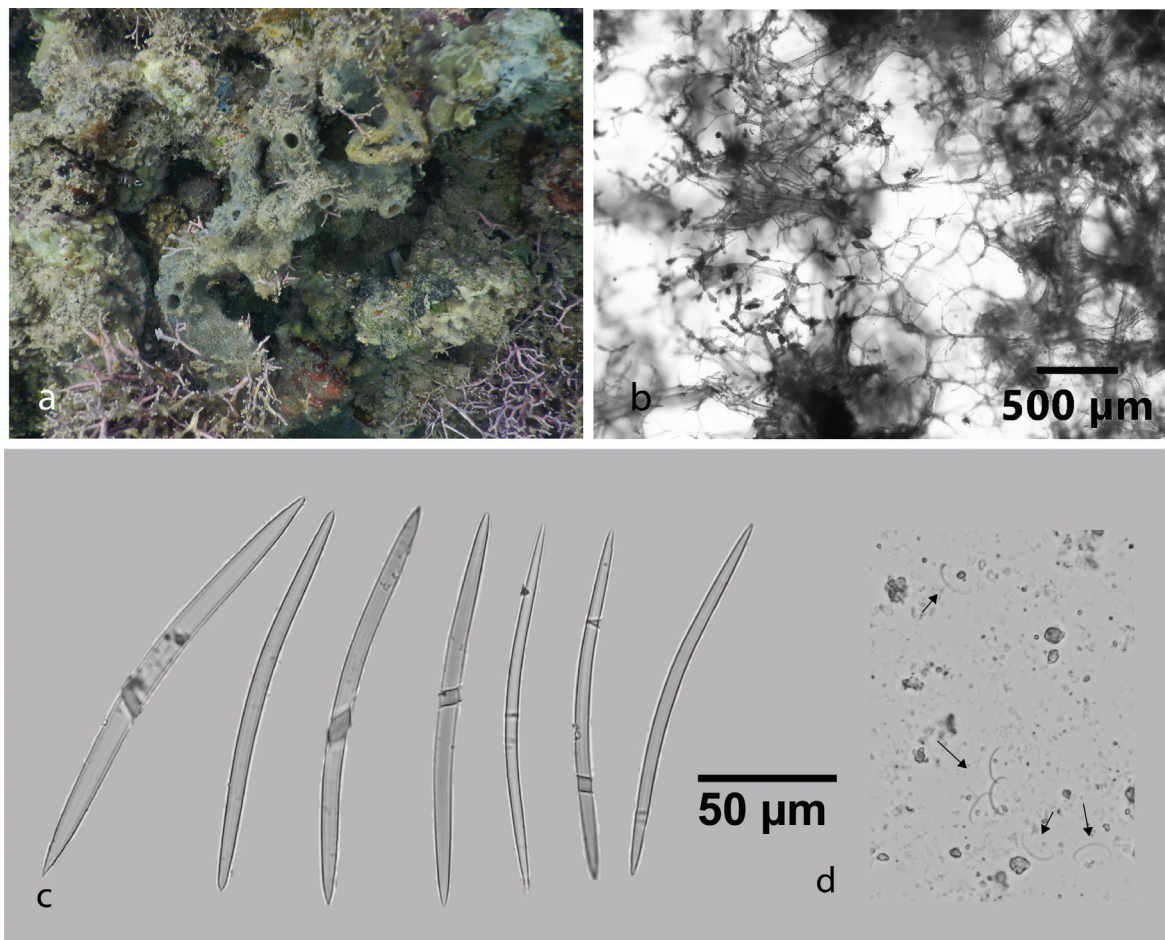


Figure 11. *Niphates nitida* Fromont, 1993 **a** habitus in situ at Kayangan Island, the Spermonde Archipelago (photograph by SAP) **b** ectosomal skeleton **c** oxeas **d** sigmas

Family Petrosiidae van Soest, 1980

Genus *Petrosia* Vosmaer, 1885

Subgenus *Petrosia* Vosmaer, 1885

***Petrosia* (*Petrosia*) *hoeksemai* de Voogd & van Soest, 2002**

Fig. 12

Diagnostic features. The sponge is thick, massive, and encrusting with rugose surface. Color brown outside, cream inside, and turning blackish brown after preservation. Choanosomal skeleton shows pauci-multispicular spicule tracts. Three sizes of oxeas, primary oxeas $182.3\text{--}272.9$ (219.6) \times $10.8\text{--}19.2$ (14.6) μm ($n = 28$), secondary oxeas $126.4\text{--}221.7$ (173.6) \times $6.7\text{--}11.4$ (8.7) μm ($n = 32$), and tertiary oxeas $58\text{--}123.9$ (83.1) \times $5.6\text{--}10.5$ (7.5) μm ($n = 28$).

Seven species of *Petrosia* have been reported from the Spermonde Archipelago, i.e., *Petrosia* (*Petrosia*) *hoeksemai* de Voogd & van Soest, 2002, *Petrosia* (*Petrosia*) *alfiani* de Voogd & van Soest, 2002, *Petrosia* (*Petrosia*) *lignosa* Wilson, 1925, *Petrosia* (*Petrosia*) *nigricans* Lindgren, 1897, *Petrosia* (*Petrosia*) *plana* Wilson, 1925, *Petrosia* (*Strongylophora*) *cortica* (Wilson, 1925),

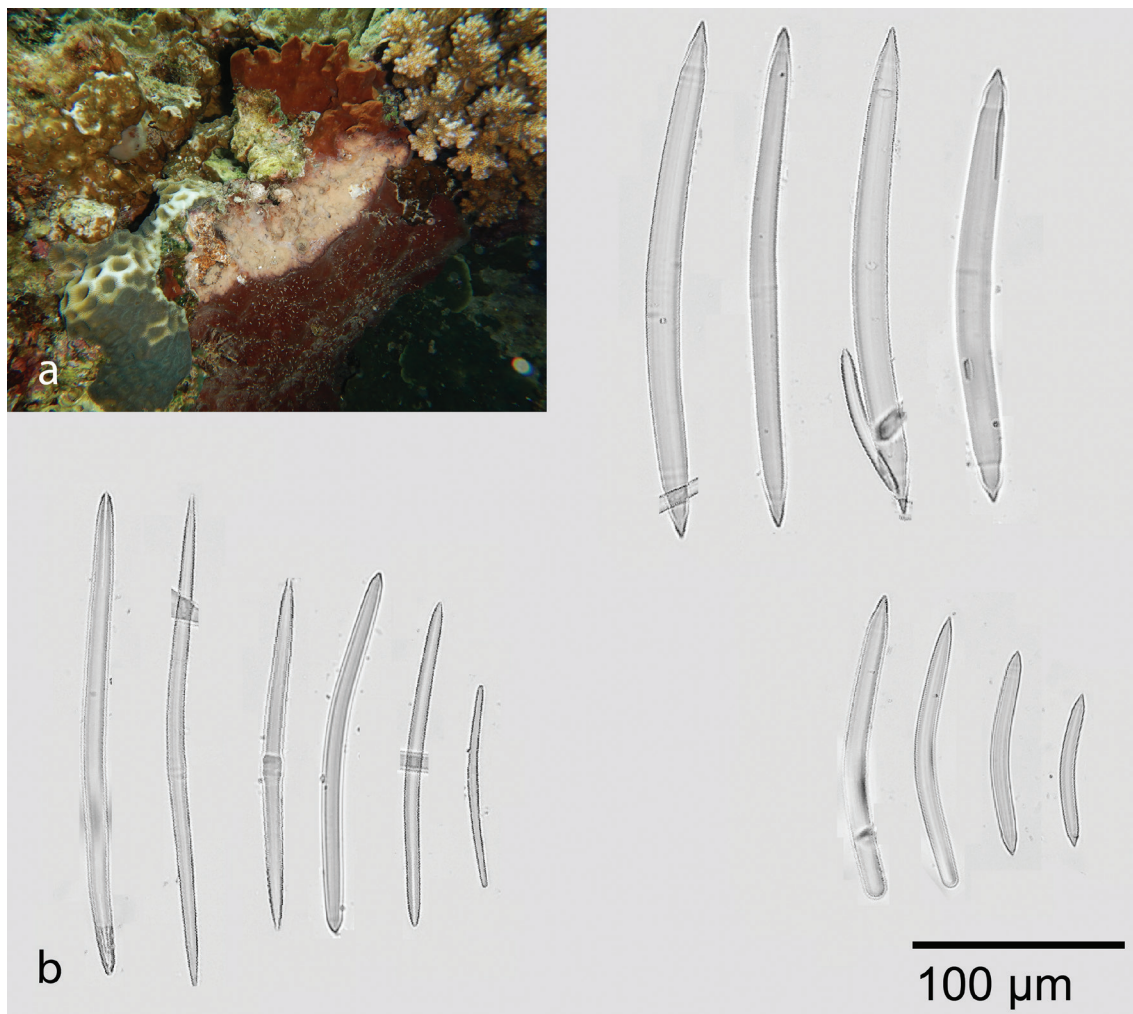


Figure 12. *Petrosia* (*Petrosia*) *hoeksemai* de Voogd & van Soest, 2002 **a** habitus in situ at Samalona Island, the Spermonde Archipelago (photograph by SAP) **b** three sizes of oxeas.

and *Petrosia (Strongylophora) strongylata* (Thiele, 1903). Two species were originally described from this area, *Petrosia (Petrosia) alfiani* and *Petrosia (Petrosia) hoeksemai* (de Voogd and van Soest 2002). Our specimen shows slightly bigger secondary and tertiary oxeas compare to the de Voogd & van Soest (2002) specimen. Comparison of spicules measurement between Indonesian *Petrosia* specimen are shown in Table 1.

Distribution and ecology. Samalona Island, the Spermonde Archipelago, attached vertically; reef flat; also reported from north Sulawesi (de Voogd and van Soest 2002).

Order Poecilosclerida Topsent, 1928

Family Coelosphaeridea Dendy, 1922

Genus *Lissodendoryx* Topsent, 1892

Subgenus *Waldoschmittia* de Laubenfels, 1936

***Lissodendoryx (Waldoschmittia) schmidti* (Ridley, 1884)**

Fig. 13

Diagnostic features. Ectosome is formed of tangentially arranged tylotes and ascending bundles in a plumose arrangement. Main skeleton is an irregular reticulation of oxeas, with triangular meshes of spicules. Microscleres are isochelas and sigmas (Hofman and van Soest 1995).

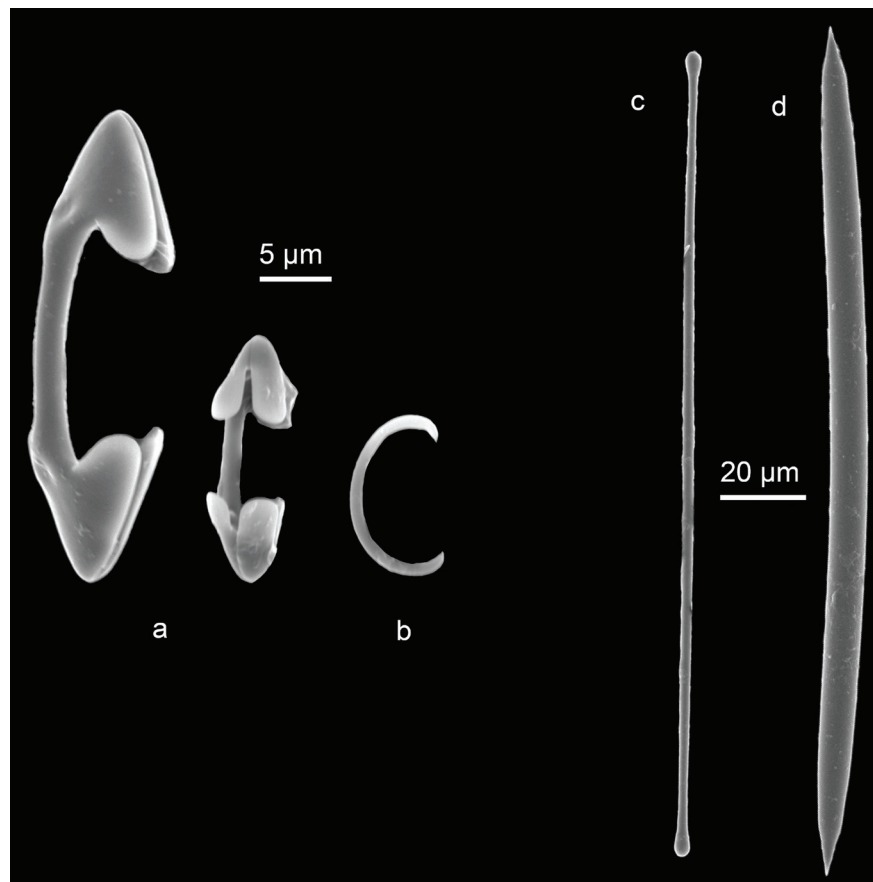


Figure 13. *Lissodendoryx (Waldoschmittia) schmidti* (Ridley, 1884) (CEL079) **a** SEM images of isochelae **b** sigma **c** tylote **d** oxea.

Distribution and ecology. This species also known from mesophotic zone. Previously recorded from Cochinchina, East Africa, Hawaii, Red Sea, Seychelles, and South Australia. In parts of Indonesia it was recorded from Ternate, Banda Sea, Aru Island (Arafura Sea), Flores, Jedan Island, East Java, and Sumba (Hofman and van Soest 1995). Our specimen is the first record for the Spermonde Archipelago, Lumulumu Island; reef flat.

Family Iotrochotidae Dendy, 1922

Genus *Iotrochota* Ridley, 1884

Iotrochota baculifera Ridley, 1884

Fig. 14

Diagnostic features. Black, thin, encrusting with rough surface, and boring. Choanosomal skeleton show multispicular reticulation. Spicule arrangements are styles 157.9–212.5 (191.7) × 7.4–15.9 (11.4) μm ($n = 25$), strongyles 248–287.6 (266.6) × 3.6–7.8 (6.7) μm ($n = 25$), with microsclere birotulate chelae, 13.9–17.3 (15.4) μm ($n = 21$). *Iotrochota baculifera* has similar coloration with *Iotrochota purpurea* (Bowerbank, 1875) and *Iotrochota nigra* (Baer, 1906). Table 2 shows the comparison of the spicule measurements of these species.

Distribution and ecology. Widespread from the Western Indian Ocean to Hawaii (Núñez Pons et al. 2017). Only two species of *Iotrochota* have been recorded from Spermonde Archipelago, *Iotrochota purpurea* and *Iotrochota baculifera*

Table 1. Comparison of spicule measurements (μm) in specimens of *Petrosia* (*Petrosia*) and *Petrosia* (*Strongylophora*) from Indonesia.

Species	Oxeas/ Strongyles 1	Oxeas/ Strongyles 2	Oxeas/ Strongyles 3	Reference
<i>Petrosia</i> (<i>Petrosia</i>) <i>hoeksemai</i>	182.3–272.9 × 10.8–19.2	126.4–221.7 × 6.7–11.4	58–123.9 × 5.6–10.5	This study
<i>Petrosia</i> (<i>Petrosia</i>) <i>hoeksemai</i>	240–305 × 10–20	90–130 × 7–12	40–75 × 5–9	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Petrosia</i>) <i>alfiani</i>	183–253 × 10–15	106–153 × 7–14	60–70 × 6–7	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Petrosia</i>) <i>lignosa</i>	230–300 × 14–18	75–150 × 10–13	35–65 × 7–10	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Petrosia</i>) <i>nigricans</i>	240–305 × 8–16	120–188 × 9–10	57–85 × 5	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Petrosia</i>) <i>plana</i>	190–290 × 7–14	95–130 × 7–9.5	43–75 × 5–9	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Strongylophora</i>) <i>cortica</i>	300–360 × 11–14	80–200 × 11–14	21–50 × 3–9	(de Voogd and van Soest 2002)
<i>Petrosia</i> (<i>Strongylophora</i>) <i>strongylata</i>	326 × 18	95–145 × 10–12	44–60 × 8–12	(de Voogd and van Soest 2002)

Table 2. Comparison of spicule measurements (μm) in specimens of *Iotrochota baculifera*, *Iotrochota purpurea*, and *Iotrochota nigra*.

Species	Styles	Strongyles	Birotulates	Reference
<i>Iotrochota baculifera</i>	157.9–212.5 (191.7) × 7.4–15.9 (11.4)	248–287.6 (266.6) × 3.6–7.8 (6.7)	13.9–17.3	This study
<i>Iotrochota baculifera</i>	200 × 9.5–12.7	220–280 × 6.3	16	(Ridley 1884)
<i>Iotrochota baculifera</i>	125–180 × 5.5–7.5	225–255 × 3.5–5	13–16.5	(Bergquist 1965)
<i>Iotrochota baculifera</i>	168–189 (175) × 4–8 (6)	201–243 (225) × 4–6 (4)	12	(Thomas 1973)
<i>Iotrochota baculifera</i>	145–170 (160) × 5–8.7 (7.5)	205–230 (220.9) × 2.5–5 (4)	12	(Núñez Pons et al. 2017)
<i>Iotrochota purpurea</i>	146–180(163) × 4–8(5)	-	16	(Thomas 1973)
<i>Iotrochota purpurea</i>	168 × 8	-	-	(Thomas 1991)
<i>Iotrochota nigra</i>	170 × 6	-	-	(Baer 1906)
<i>Iotrochota nigra</i>	230–269 (251) × 5 (5)	163–193(184) × 7(7)	17(17)	(Samaai et al. 2019)

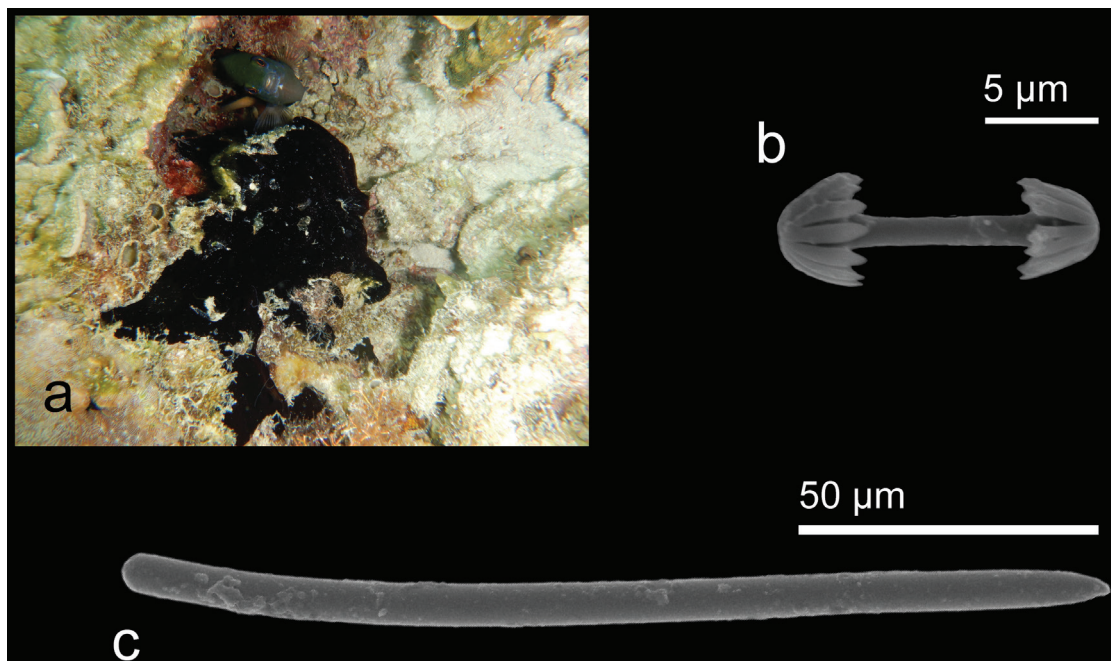


Figure 14. *Iotrochota baculifera* Ridley, 1884 a habitus in situ at Samalona Island, the Spermonde Archipelago (photograph by SAP) b birotulate chelae c styles.

Ridley, 1884 (de Voogd 2005). Our specimen was found in the north-west of Samalona Island, the Spermonde Archipelago; reef flat.

Family Microcionidae Carter, 1875

Genus *Clathria* Schmidt, 1862

Subgenus *Thalysias* Duchassaing & Michelotti, 1864

***Clathria (Thalysias) reinwardti* Vosmaer, 1880**

Fig. 15

Diagnostic features. Arborescent, simple massive, and very repent appearance with many small oscula. Bright to dark orange in living material, and brown in alcohol. Reticulate skeleton with two class sizes of styles and echinating acanthostyles. Principal styles slightly curved with strongylote point, 151–312 (205.5) × 5.3–10.85 (7.4) µm ($n = 28$), auxiliary styles straight and slightly curved, 72–163 (106.5) × 1.5–4.7 (3.4) µm ($n = 37$), and echinating acanthostyles with short, rounded point and dense spines on point and base, 51.9–81.5 (67.1) × 6.2–8.7 (7.3) µm ($n = 31$). This species can be differentiated from other similar *Thalysias* by its characteristic acanthostyle morphology, growth form, and the size and geometry of its toxas, including ectosomal-subectosomal features (Hooper 1996). Hooper's (1996) specimen shows microscleres as palmate isochelae in two size classes and oxhorn toxas.

Distribution and ecology. Central Indian Ocean (Thomas 1986), Indo-Pacific (van Soest 1990; Lim et al. 2016), and Australia (Hooper 1996). Commonly found in coral rubble or dead coral and hard substrates. Our specimen was found in the Spermonde Archipelago, the north-west of Samalona Island; reef flat and Gusung Tallang; turbid reef.

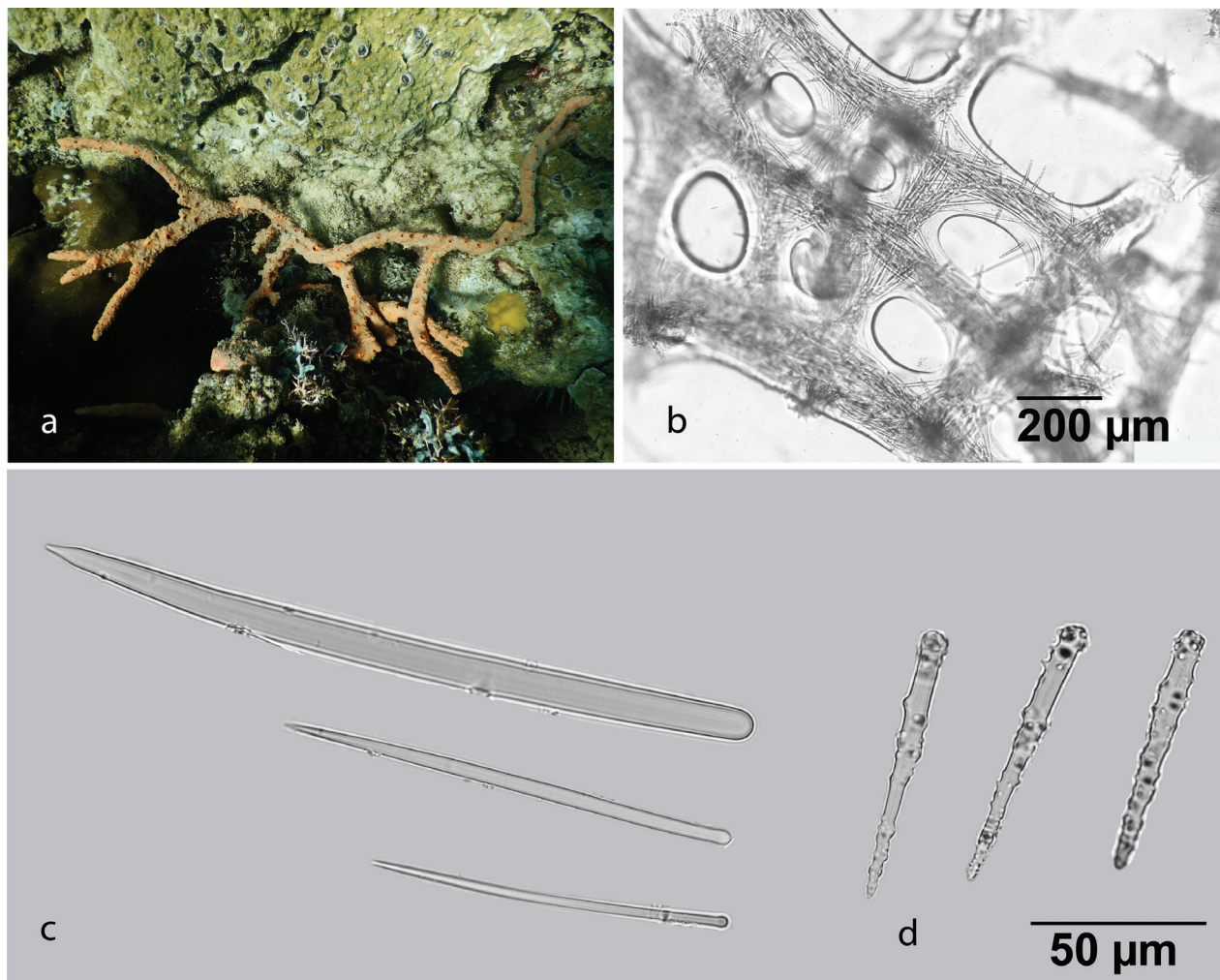


Figure 15. *Clathria (Thalysias) reinwardti* Vosmaer, 1880 **a** habitus in situ at Samalona Island, the Spermonde Archipelago (photograph by SAP) **b** longitudinal section of the skeleton **c** styles **d** acanthostyles.

Order Scopalinida Morrow & Cárdenas, 2015

Family Scopalinidae Morrow et al., 2012

Genus *Stylissa* Hallmann, 1914

***Stylissa massa* (Carter, 1887)**

Fig. 16

Diagnostic features. Massive, soft, and friable with rough surface and medium-sized oscula appear on top of the ridge. Yellow-orange in life and brown in alcohol. Spicules arrangements are of styles and strongyles.

Distribution and ecology. *Stylissa massa* is widely distributed in the Indo-Pacific (Erpenbeck et al. 2017). Since *Stylissa massa* is known to be widespread, and recent studies using molecular techniques show the probability of distinct cryptic lineages of this species in the Indo-Pacific (Erpenbeck et al. 2017). Our specimen was collected from the Spermonde Archipelago, south-west of Samalona Island; reef flat, attached to rubble and dead coral skeletons.



Figure 16. Habitus in situ of *Stylissa massa* (Carter, 1887) at Samalona Island, the Spermonde Archipelago (photograph by SAP).

Order Suberitida Chombard & Boury-Esnault, 1999

Family Halichondriidae Gray, 1867

Genus *Halichondria* Fleming, 1828

Subgenus *Halichondria* Fleming, 1828

***Halichondria (Halichondria) cartilaginea* (Esper, 1797)**

Fig. 17

Description. Massive creeping growth form with upright branches (branching). These branches are irregular and form mats covering the substrate. Color bright green, flexible/cartilaginous. This species lives in association with Chlorophyta *Cladophoropsis vaucheriiformis* (Areschoug) Papenfuss, 1958 (van Soest 1990). Spicules are only oxeas, 125.46–252.45 (191.40) × 4.03–7.40 (5.37) μm.

Distribution and ecology. Currently this species is recorded from China, Vietnam, Malacca Strait, Banda Sea, and East African Coral Coast. According to the WPD checklist (de Voogd et al. 2024), this is the first record from the Spermonde Archipelago, Badi Island; reef flat.

Genus *Topsentia* Berg, 1899

***Topsentia indica* Hentschel, 1912**

Fig. 18

Description. Only two species of *Topsentia* are distributed in Indonesia, i.e., *Topsentia dura* (Lindgren, 1897) and *Topsentia indica* Hentschel, 1912. *Topsentia dura* had further illustrations and spicule measurements provided



Figure 17. Habitus in situ of *Halichondria (Halichondria) cartilaginea* (Esper, 1797) (CEL025) (photograph by NJdeV).

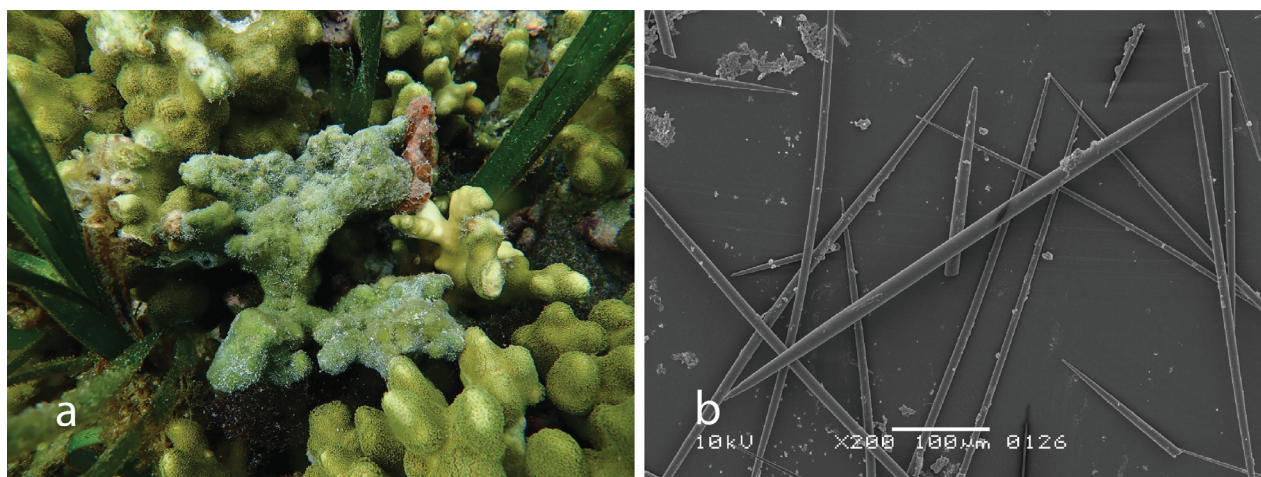


Figure 18. *Topsentia indica* Hentschel, 1912 a habitus in situ at Langkai Island, the Spermonde Archipelago (photograph by NJdeV) b SEM images of the spicules.

by a previous study (Alvarez and Hooper 2011). These species are massive, of hard consistency with skeletons made of a confused mass of oxeas of similar dimensions, not clearly differentiated into size classes. Our specimen shows similar characteristics with the specimen of Hentschel (1912).

Distribution and ecology. Previously recorded from Aru Islands (Hentschel 1912). This is first record from the Spermonde Archipelago, Langkai Island; reef flat.

Family Suberitidae Schmidt, 1870

Genus *Suberites* Nardo, 1833

***Suberites* sp.**

Fig. 19

Diagnostic features. Ficiform (fig-shaped) with orange (almost red) color and fragile. Oscula found on top of the fig-like shape. Aquiferous network can be seen from ectosomal skeleton of living specimen, small ostia also visible. Spicules are tylostyles (total length \times width) 204.3–324.5 (278.4) \times 3.5–8.6 (5.5) μm ($n = 31$). Tylostyle heads are oval with an indistinct neck (head length \times head width \times neck width) 8.8–15.9 (12.3) \times 4.2–8.8 (6) \times 3.2–8 (4.5) μm ($n = 25$).

Distribution and ecology. Only three *Suberites* species have been recorded from Indonesia, *Suberites radiatus* Kieschnick, 1896, *Suberites diversicolor* Becking & Lim, 2009, and the deep-sea *Suberites perfectus* Ridley & Dendy, 1886 (Becking and Lim 2009; Putra et al. 2023). North-west of Samalona Island, the Spermonde Archipelago; reef flat, scattered across shallow water area, growth on rock, plastic PVC, and sometimes competing with Scleractinia.

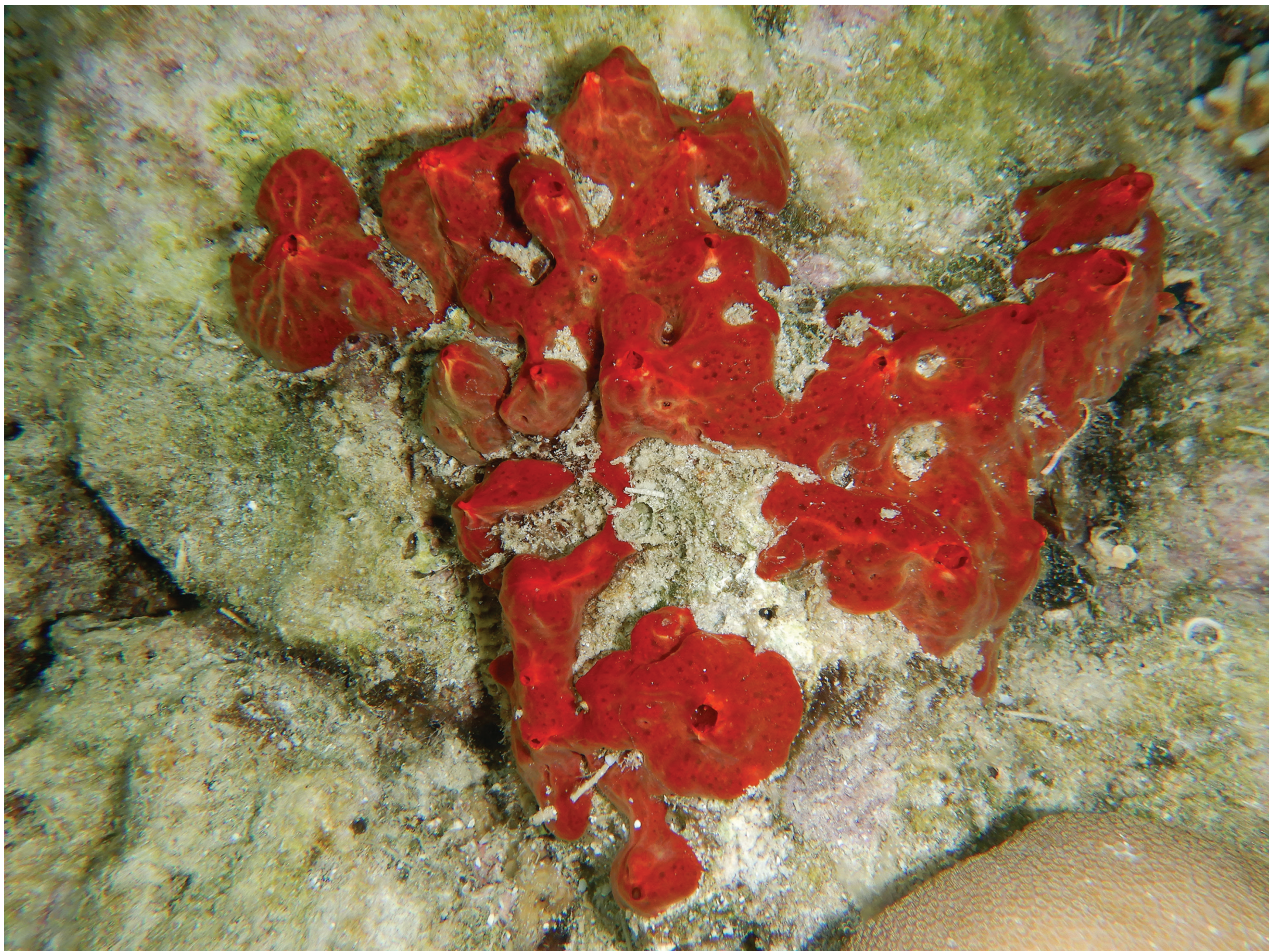


Figure 19. Habitus in situ of *Suberites* sp. (photograph by SAP).

Genus *Terpios* Duchassaing & Michelotti, 1864

***Terpios hoshinota* Rützler & Muzik, 1993**

Fig. 20

Diagnostic features. Thin (< 1 mm thick), encrusting, and excavating form overgrowing host coral skeletons (*Acropora* spp.). Dark grey to black, sometime pale grey in the upper surface. Original description of *Terpios hoshinota* show spicules as only tylostyles (Rützler and Muzik 1993). In this study, spicule arrangements are tylostyles (total length × width) 132.9–252 (206.9) × 2.6–7.8 (4.4) μm ($n = 52$), and variation of heads (head length × head width × neck width) 3.7–7.4 (5.4) × 4.8–9 (6.5) × 1.8–5 (3.3) μm ($n = 27$). Spicule dimension measurements are shown on Table 3. The morphology of *Terpios hoshinota* is similar to *Terpios granulosus* Bergquist, 1967 from Hawaiian reefs. The difference is that this species is greyish brown, has lobe-headed tylostyles, and has a cyanobacterial symbiont (Rützler and Muzik 1993). This species known as a coral-killing sponge, but a recent study shows *Terpios hoshinota* could also grow on glass slides, plastic sheets, and rubber tyres. The competitive interaction with the coral host is only for substrate rather than food or nutrients (Syue et al. 2021).

Distribution and ecology. This widespread species has been recorded from the Indian Ocean, north-western Pacific, and Australia (Fromont et al. 2019).

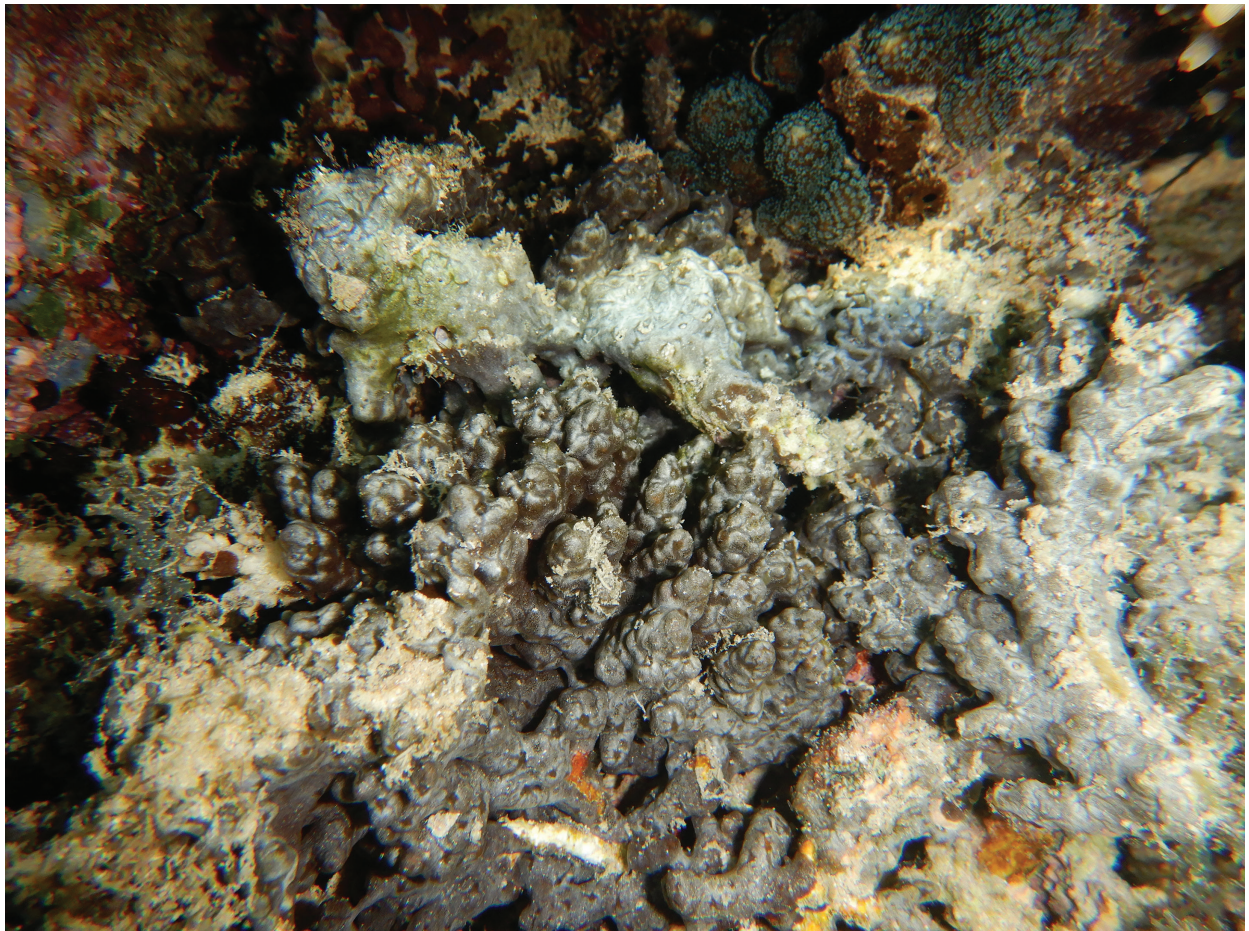


Figure 20. Habitus in situ of *Terpios hoshinota* Rützler & Muzik, 1993 (photograph by SAP).

Table 3. Spicule (tylostyles) dimensions (μm) for *Terpios hoshinota*.

Total length	Shaft width	Neck width	Head width	Head length	Reference
132.9–252 (206.9)	2.6–7.8 (4.4)	1.8–5 (3.3)	4.8–9 (6.5)	3.7–7.4 (5.4)	This study
180–290 (251.6)	3–4 (3.5)	2–3 (2.7)	5.5–7 (6.1)	4.5–6 (5.2)	(Rützler and Muzik 1993)

Terpios hoshinota was originally described from the Ryukyu Archipelago, Japan (north-west Pacific). Our specimen was found from north-west of Samalona Island, the Spermonde Archipelago; reef flat, overgrowing branching *Acropora* sp.

Order Tetractinellida Marshall, 1876

Family Ancorinidae Schmidt, 1870

Genus *Ecionemia* Bowerbank, 1862

***Ecionemia acervus* Bowerbank, 1862**

Fig. 21

Description. Massive or thickly encrusting sponges without a distinct cortex. Megascleres are triaenes of different types and large oxeas. Microscleres in-

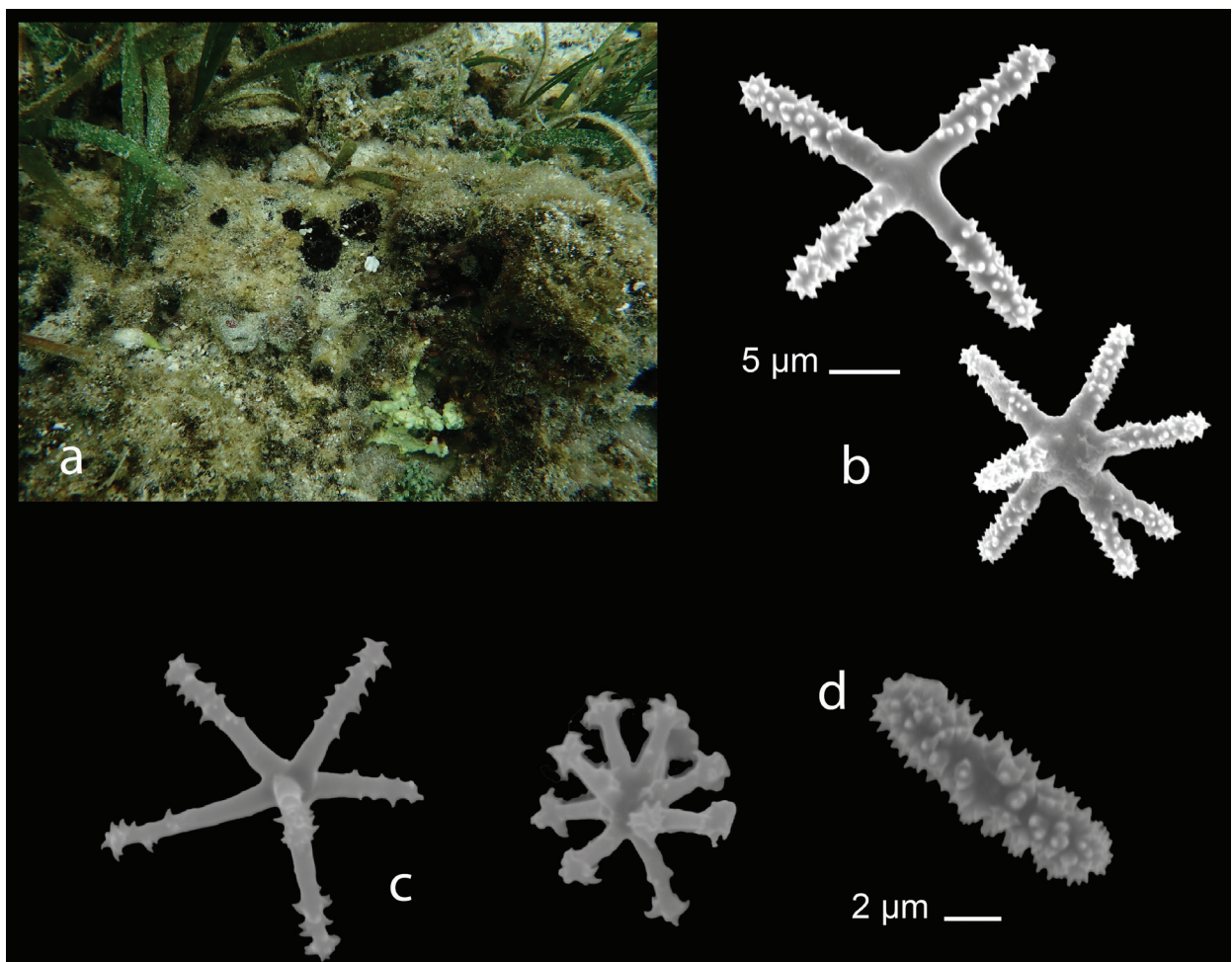


Figure 21. *Ecionemia acervus* Bowerbank, 1862 **a** habitus in situ at Langkai Island, the Spermonde Archipelago (photograph by NJdeV) (CEL016), and SEM images of spicules, **b**, **c** somal chiasters/ strongylasters **d** cortical rough microrhabds/ microstrongyle.

clude spiny microrhabds in addition to euasters. Microrhabds usually form a dermal layer (Uriz 2002).

Distribution and ecology. Indo-Pacific, Australia, New Zealand. This species is common in the Indo-Pacific (Uriz 2002). Our specimen was collected from Langkai Island, the Spermonde Archipelago; reef flat.

Family Geodiidae Gray, 1867

Genus *Geodia* Lamarck, 1815

***Geodia* sp.**

Fig. 22

Diagnostic features. Twelve species of *Geodia* spp. were described from Indonesia (Putra et al. 2023; de Voogd et al. 2024). Our specimen has oxeas 1079.43–1820.54 (1507.20) × 18.17–33.67 (25.21) μm (*n* = 11), sterrasters widths 49.05–77.40 (59.98) μm (*n* = 20), dichotriaene, anatriaene, protriaene, strongylasters, and oxyasters. Further analysis is needed to examine and provide a name for this specimen.

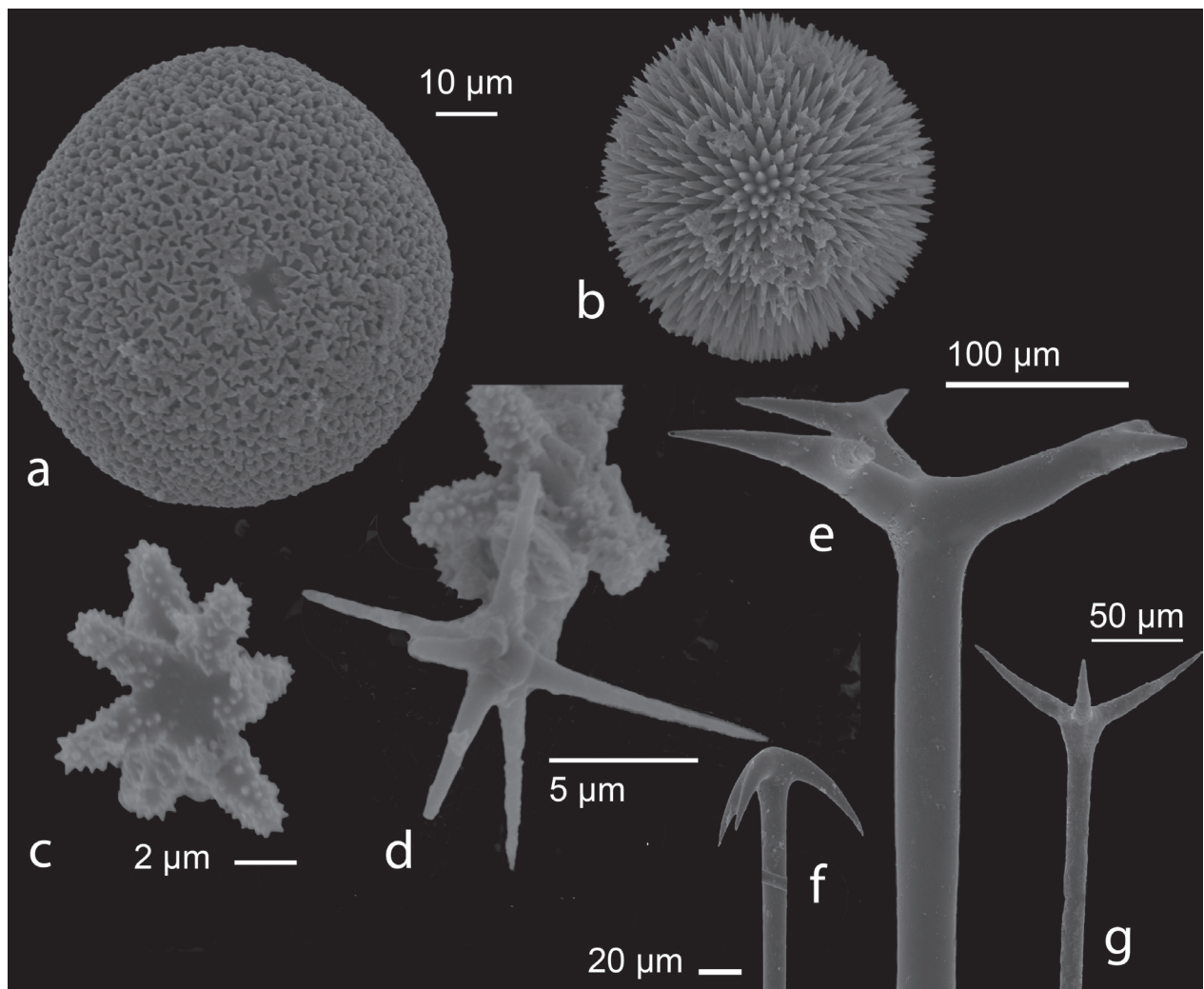


Figure 22. SEM images of spicules of *Geodia* sp. (CEL174) **a**, **b** sterrasters (**b** developmental stage) **c** strongylaster **d** oxyaster attached to strongylasters **e** dichotriaene **f** anatriaene **g** protriaene.

Distribution and ecology. This group is distributed across Indonesia, i.e., Halmahera, Arafura Sea, Southern Java, Sunda Shelf/Java Sea, Banda Sea, Palawan/North Borneo (Sollas 1888; Kieschnick 1896; Topsent 1897; Lindgren 1898; Thiele 1900; von Lendenfeld 1903; Hentschel 1912; Wilson 1925; van Soest et al. 2020). Our specimen was collected from Barangbaringan Island, the Spermonde Archipelago; turbid environment.

Family Tetillidae Sollas, 1886
Genus *Paratetilla* Dendy, 1905

***Paratetilla bacca* (Selenka, 1867)**

Fig. 23

Diagnostic features. Globular sponges, specimen $\approx 64 \times 47$ mm (l \times w) in diameter. Porocalices are abundant as circular to oval apertures. Color generally bright yellow when alive with brownish appearance in situ due to algal and sediment cover. Skeleton composed of oxea and triaenes radiating from a central core. Megascleres are oxeas, anatriaenes, and calthrops-like. Microscleres are sigmaspires, C- to S-shaped. A complete redescription of *P. bacca* was provided recently (Santodomingo and Becking 2018). This species had a considerable variation in spicules sizes in the different localities as well as significant intra-specific variation. This variation could be a response to different environmental conditions, a consequence of genetic selection, or synergistic between ecological and genetic factors.

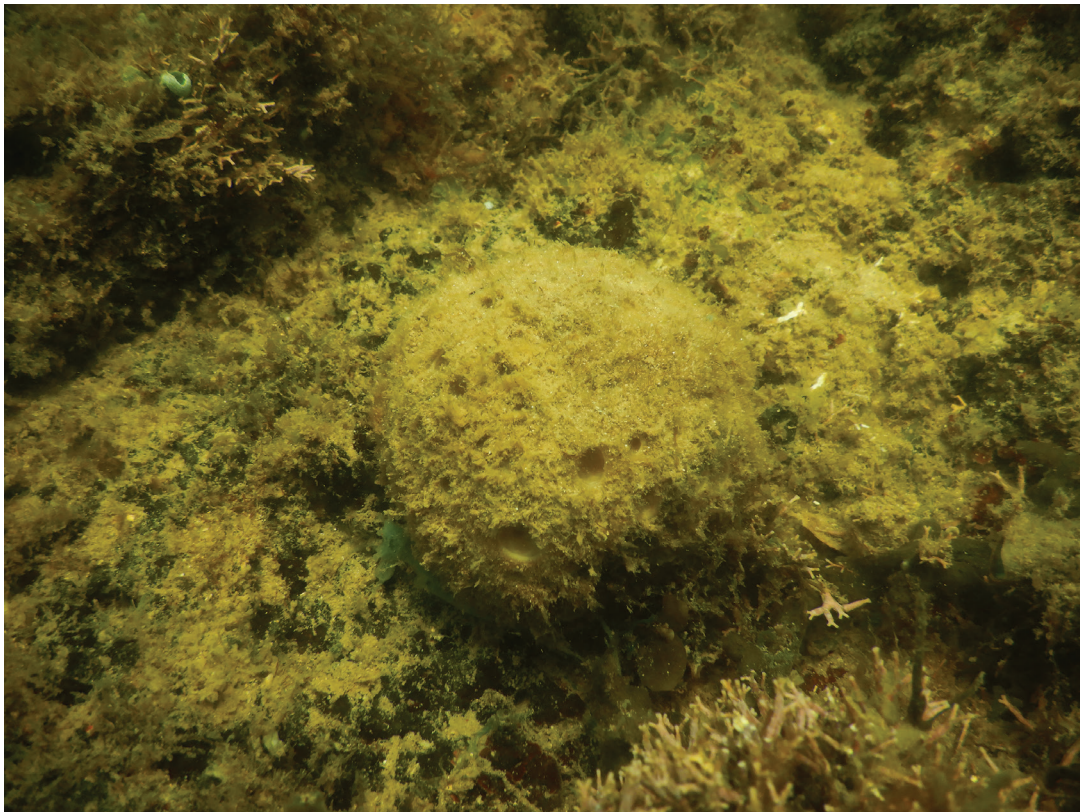


Figure 23. Habitus in situ of *Paratetilla bacca* (Selenka, 1867) at Gusung Tallang, the Spermonde Archipelago (photograph by SAP).

Distribution and ecology. Seychelles Islands (Thomas 1973), south-west Madagascar (Vacelet et al. 1976), Zanzibar (Pulitzer-Finali 1993), Thailand (Putchar-karn 2007), Singapore (Lim et al. 2012), Philippines (Longakit et al. 2005), and Indonesia (Santodomingo and Becking 2018). Our specimen was collected from Gusung Tallang Island, the Spermonde Archipelago; turbid environment.

Subclass Keratosa Grant, 1861

Order Dictyoceratida Minchin, 1900

Family Dysideidae Gray, 1867

Genus *Lamellodysidea* Cook & Bergquist, 2002

***Lamellodysidea herbacea* (Keller, 1889)**

Fig. 24

Diagnostic features. Live specimen found was white to pale green in color, and grey after preservation. This species habitus is soft, fragile, slick, thin (< 1 cm thick), and has an encrusting basal plate with a complex labyrinthine wall-like pattern. Skeleton structure forming interconnected reticulate fibers with several adjacent spicules. Various of microsymbionts (cyanobacteria) are found inhabiting it. Currently there only two species of *Lamellodysidea*, *Lamellodysidea herbacea* (Keller, 1889) and *Lamellodysidea chlorea* (de Laubenfels, 1954), both confused with each other. *Lamellodysidea herbacea* is known to be com-



Figure 24. Habitus in situ of *Lamellodysidea herbacea* (Keller, 1889) at Samalona Island, the Spermonde Archipelago (photograph by SAP).

mon in the sub-intertidal zone of the coral reef, which is exposed to sunlight (Putchakarn 2007). Molecular analysis shows *Lamellodysidea herbacea* is a diverse group and consists of several distinct lineages of the alleged single species, and has probably been misidentified in the past with undescribed lineages due to superficial resemblances (Erpenbeck et al. 2012).

Distribution and ecology. Our specimen was collected from Samalona Island, the Spermonde Archipelago; reef flat. This species was previously recorded from the Red Sea (Row 1911), India (George et al. 2020), Thailand (Putchakarn 2007), the Spermonde Archipelago (de Voogd et al. 2006), and the Great Barrier Reef (Hooper 2008).

Family Irciniidae Gray, 1867

Genus *Ircinia* Nardo, 1833

***Ircinia schulzei* (Dendy, 1905)**

Fig. 25

Diagnostic features. Specimen attached to hard substrate, cylindrical with irregular short or club-shaped branches and rugose surface. Color in life is pale green and pale white in alcohol. Small oscula are found in every branch, sometimes on the tip. Skeleton is laminated fiber. Irciniidae are massive, or occasionally encrusting, sponges that display a wide range of forms, e.g., caliculate, lamelliform, lobate, and digitate. The species of *Ircinia* are pithed and laminated with primary and secondary fibers (de C. Cook and Bergquist 1999).

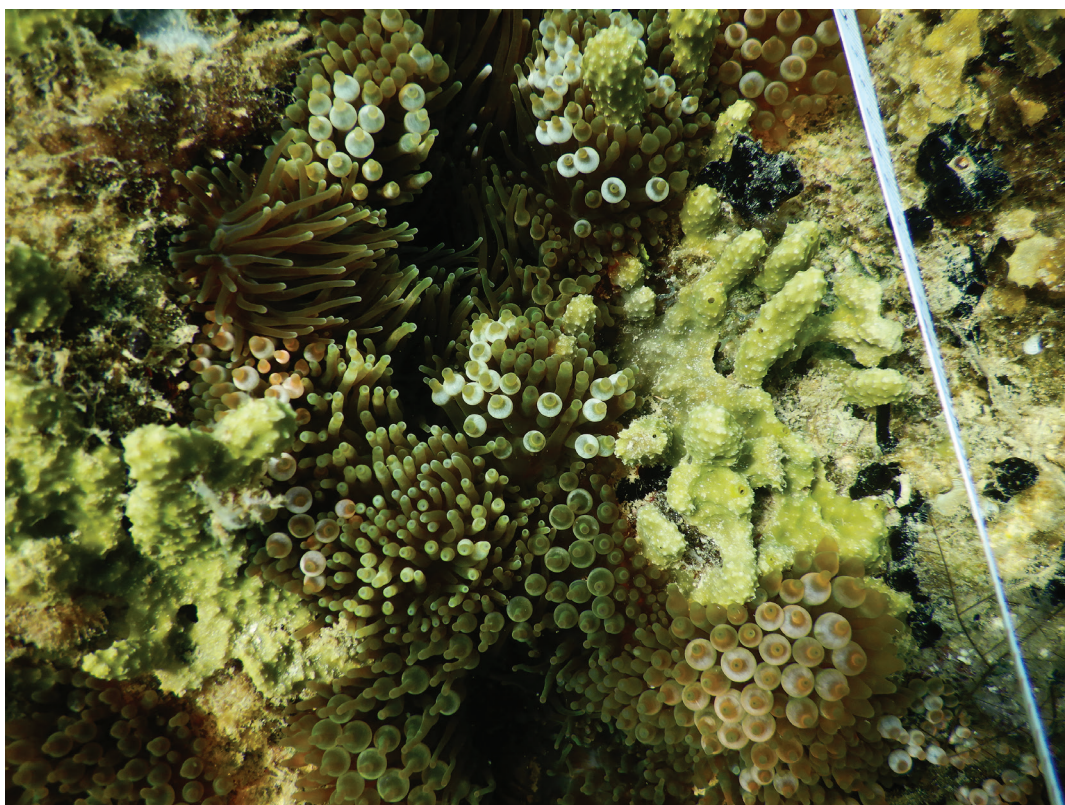


Figure 25. Habitus in situ of *Ircinia schulzei* (Dendy, 1905) at Samalona Island, the Spermonde Archipelago (photograph by SAP).

Distribution and ecology. *Ircinia schulzei* was first described from Ceylon (Sri Lanka today; Dendy 1905). A previous record from Papua New Guinea (Pulitzer-Finali and Pronzato 1999) and this new record in the Spermonde Archipelago shows that the species could be widespread in the Indo-Pacific region. Our specimen was found living between an anemone and other sponges on top of a rock in the reef flat of north-west of Samalona Island, the Spermonde Archipelago.

Family Thorectidae Bergquist, 1978

Genus *Phyllospongia* Ehlers, 1870

***Phyllospongia foliascens* (Pallas, 1766)**

Fig. 26

Diagnostic features. Specimen form is foliaceous and irregular flabellate branches, pale white color in life and when preserved, < 0.5 mm thick. Numerous small oscula (< 1 mm) scattered in the surface. Skeleton consists of interconnected reticulate fibers. This species was recently transferred from the genus *Carteriospongia* Hyatt, 1877 due to molecular phylogenetic analysis showing *Carteriospongia foliascens* as a clade member of *Phyllospongia bergquistae* Abdul Wahab & Fromont, 2020. The original diagnosis describing a verrucose surface is characteristic for *Phyllospongia foliascens*, but with a fine and meandering surface patterning for *Phyllospongia bergquistae* (Bergquist et

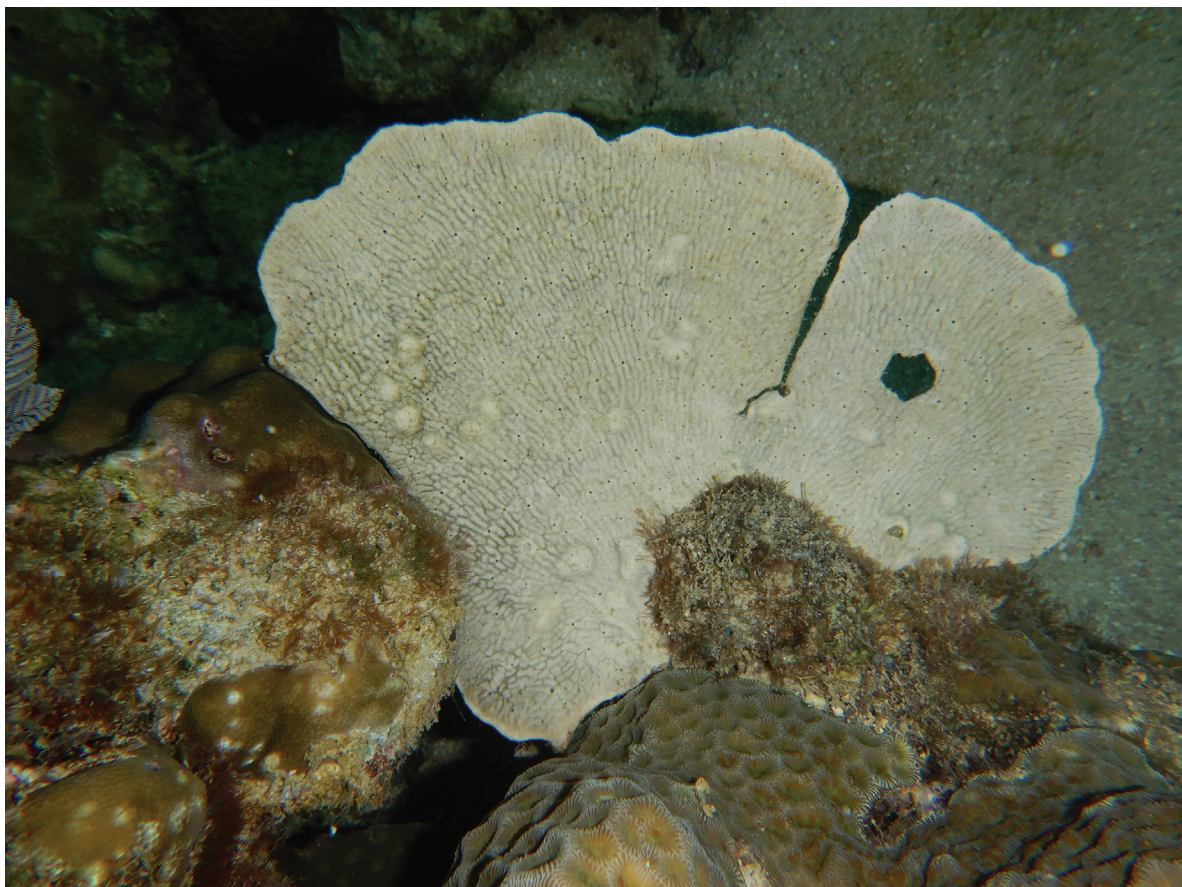


Figure 26. Habitus in situ of *Phyllospongia foliascens* (Pallas, 1766) at Samalona Island, the Spermonde Archipelago (photograph by SAP).

al. 1988; Abdul Wahab et al. 2021). *Phyllospongia foliascens* is a phototrophic species that mainly relies on symbiotic cyanobacteria for its nutrient cycle. This species is also able to endure high energy environments (Cleary et al. 2005).

Distribution and ecology. Numerous individuals were found during the survey. *Phyllospongia foliascens* is widely distributed and has a high density in the Spermonde Archipelago (de Voogd et al. 2006). Our specimen was found at south-west of Samalona Island, reef flat; Gusung Tallang, turbid reef. This species has been recorded from shallow waters of the Red Sea, Indian Ocean, Australia, and Fiji (Abdul Wahab et al. 2021).

Discussion

Twenty-seven species of marine sponges (Class Calcarea and Class Demospongiae) have been identified in the littoral area of the Spermonde Archipelago, Indonesia. The Order Haplosclerida, with nine species, dominates this type of habitat. According to the WPD checklist (de Voogd et al. 2024), some of the sponges found here, such as *Clathrina rodriguesensis*, *Amphimedon paraviridis*, *Niphates nitida*, and *Ircinia schulzei*, are newly recorded for Indonesia. Several others are new records for the Sulawesi Sea/Makassar Strait marine ecoregion, including *Janusya tubuloreticulosa*, *Leucaltis nodusgordii*, *Spirastrella* aff. *decumbens*, *Haliclona (Reniera) venusta*, *Haliclona (Soestella) elegantia*, *Lissodendoryx (Waldoschmittia) schmidtii*, *Halichondria (Halichondria) cartilaginea*, and *Topsentia indica* (Suppl. material 2). Four species potentially new to science are also preliminarily described; further examination, including molecular analysis, is needed to accurately describe all the species.

In relation to extreme habitats, several species such as *Phyllospongia foliascens*, *Stylissa massa*, *Clathria (Thalysias) reinwardti*, and *Haliclona (Gellius) cymaeformis* are frequently found in this habitat (Suppl. material 3). For instance, the foliose sponge *Phyllospongia foliascens* as well as *Haliclona (Gellius) cymaeformis* were very abundant in the turbid reef near Makassar city, e.g., Kayangan Island, Gusung Tallang, and Samalona Island (SAP pers. obs. 2020). This habitat is unusual for phototrophic species. Studies in other areas (i.e., north-west Java, the Great Barrier Reef) have shown that they are typically found in oligotrophic environments, characterized by low concentrations of organic nutrients (Wilkinson 1988; de Voogd and Cleary 2008). Conversely, several variables could be influencing the presence of these species in this unique environment. This could also be altered by algal symbionts that provide all the required carbon through photosynthesis, and the nitrogen from heterotrophic sources such as ultra-plankton (Davy et al. 2002; Pile et al. 2003).

Several species mentioned above, including *Paratetilla bacca*, *Spirastrella decumbens*, and *Petrosia (Petrosia) hoeksemai*, have demonstrated preferences for sedimented environments (Putchakarn 2011; Schönberg 2021). Although psammobiotic species typically exhibit an affinity for sedimented habitats (Schönberg 2016), sediment presence can exert negative pressures on sponge communities. Specifically, when subjected to elevated concentrations of suspended sediment, sponge taxa can exhibit diminished pumping activity and reduced feeding efficiency (Lohrer et al. 2006). Moreover, there may be alterations in their respiration rates (Pineda et al. 2017) and tissue abrasion (Nava and Carballo 2013). Such physiological stressors can culminate in partial mortality

and compromised survival rates. A decline in sponge abundance, biomass, and species diversity has the potential to instigate cascading effects on broader marine ecosystems (Bell 2008).

Conclusions

In the littoral area, sponges predominantly colonize coral matrices and other hard substrates. Our recent investigation uncovers previously undocumented occurrences, including potentially new taxa, within the sponge community residing in the Sulawesi Sea/Makassar Strait marine ecoregion, particularly at the Spermonde Archipelago, SW Sulawesi. Noteworthy findings include the identification of 15 new records for the marine ecoregion, bringing the total to 143 species on the checklist, not including four potentially novel species. The sponge assemblage within this archipelago presents a rich and intricate biodiversity, underscoring an immediate imperative for comprehensive characterization. Rigorous examination coupled with molecular analysis of specimens is essential to ensure description of the entire species set.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

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Author contributions

Conceptualization: SAAP, JJ. Data curation: NJJV. Formal analysis: SAAP. Funding acquisition: RAR. Investigation: SAAP. Methodology: RAR. Resources: RAR. Supervision: JJ, RAR, NJJV. Validation: NJJV. Writing - original draft: SAAP. Writing - review and editing: RAR, NJJV, JJ.

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Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

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Supplementary material 1

Sampling sites of sponge (Porifera) collections from shallow-subtidal habitat of the Spermonde Archipelago, Indonesia

Authors: Singgih Afifa Putra, Rohani Ambo-Rappe, Jamaluddin Jompa, Nicole J. de Voogd

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Supplementary material 2

checklist of Porifera from Sulawesi Sea/Makassar Strait marine ecoregion with updates based on the current study

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Supplementary material 3

List of sponge (Porifera) species examined in this study with locations and environmental condition in the Spermonde Archipelago, Indonesia

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